#### Royal Entomological Society



## HANDBOOKS FOR THE IDENTIFICATION OF BRITISH INSECTS

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# HANDBOOKS FOR THE IDENTIFICATION OF BRITISH INSECTS



### COLEOPTERA INTRODUCTION AND KEYS TO FAMILIES

By R. A. CROWSON

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#### HANDBOOKS FOR THE IDENTIFICATION OF BRITISH INSECTS

The aim of this series of publications is to provide illustrated keys to the whole of the British Insects (in so far as this is possible), in ten volumes, as follows:

Part 9. Ephemeroptera.

,, 10. Odonata. ,, 11. Thysanoptera.

" 12. Neuroptera.

,, 14. Trichoptera. ,, 15. Strepsiptera.

" 16. Siphonaptera.

" 13. Mecoptera.

I. Part 1. General Introduction.

" 2. Thysanura.

" 3. Protura.

4. Collembola.

, 5. Dermaptera and Orthoptera.

, 6. Plecoptera.

" 7. Psocoptera.

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II. Hemiptera.
III. Lepidoptera.

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VI. Hymenoptera: Symphyta and Aculeata.

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Volumes II to X will be divided into parts of convenient size, but it is not possible to specify in advance the taxonomic content of each part.

Conciseness and cheapness are main objectives in this new series, and each part will be the work of a specialist, or of a group of specialists. Although much of the work will be based on existing published keys, suitably adapted, it is expected that it will also include much new and original matter.

Parts will be issued, separately paged and priced, as they become

available.

Orders for the Series or for separate parts should be sent to the Registrar at the Society's rooms.

The Society is indebted to the Royal Society for a grant towards the cost of initiating this series of *Handbooks*.

A list of parts now available appears on the back cover.

#### COLEOPTERA

#### GENERAL INTRODUCTION AND KEYS TO FAMILIES.

#### By R. A. Crowson.

The Coleoptera of the British Islands have been treated in a series of comprehensive works, the most important of which are Stephens' Manual of British Coleoptera (1839), Cox's Handbook of Coleoptera (1874), Fowler's great Coleoptera of the British Islands (1887–1891), and Joy's Practical Handbook of British Beetles (1932). Numerous British works of more limited scope have been published. Similar works on the Coleoptera of adjacent areas, including almost all the British species, are Reitter's Fauna Germanica: Käfer (1908–1916) and Portevin's Histoire Naturelle des Coléoptères de France (1929); the Dutch and Danish beetle faunas have also been the subject of systematic works. The French and German works, like the British, are now out of print and more or less difficult to obtain, and even the most recent of them are more than twenty years old. The last twenty years have seen some important advances in our knowledge of the relationships of the groups of Coleoptera, and some not inconsiderable additions to our knowledge of the beetle fauna of Great Britain.

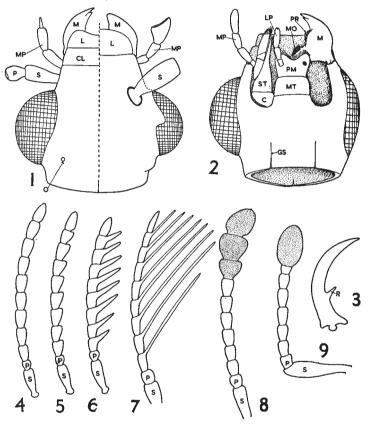
#### GENERAL CHARACTERS OF COLEOPTERA.

The Coleoptera may be defined as insects with a complete metamorphosis (Endopterygota), having the mesothoracic wings modified into hard elytra or into haltere-like structures, the metathoracic wings alone providing propulsion in flight; the mouth-parts are of the biting type and the head has a distinct gular region below. Coleopterous larvae are of the oligopod or apodous types and lack labial (spinning or salivary) glands; beetle pupae are usually of the exarate type and they never possess true functional mandibles. There are reasons for believing that the Coleoptera are most nearly related to the Megaloptera (particularly the Sialoidea) and rather less closely to the Neuroptera proper (Planipennia).

#### STRUCTURE OF ADULT COLEOPTERA.

(1) The Head (figs. 1-9).—The labrum (fig. 1 l) may be freely articulated, more or less fused to the clypeus, or reduced and hidden under the front margin of the clypeus. The clypeus (fig. 1 cl) may be separated from the frons by a complete transverse groove (fronto-clypeal suture), or the groove may be incomplete, indistinct, or absent. The frons is normally not separated by a distinct suture from the vertex, which may occasionally bear one or two occili (fig. 1 o); the posterior or occipital part of the vertex may exhibit a median longitudinal suture (usually only visible when the head is

pulled out of the prothorax), which occasionally may divide anteriorly to form a pair of widely diverging sutures ("frontal sutures" of some authors) extending towards the antennal insertions—simulating, but probably not truly homologous with, the "ecdysial suture" of larvae. The head capsule may or may not be constricted behind the eyes to form a neck (fig. 1); its posterior part is invariably more or less retracted into the prothorax. The

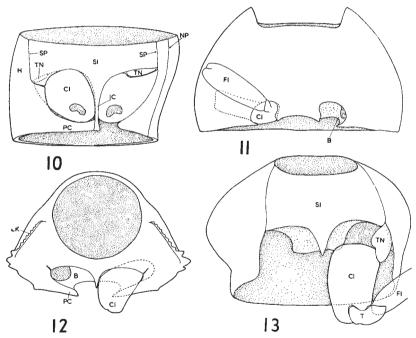


Figs. 1-9.—Head structures in Coleoptera. 1, Composite head in dorsal view: left side showing entire eye, aciculate maxillary palp (MP), apically bifid mandible, distinct clypeus, antennae inserted under sides of front; right side showing head capsule contracted behind eye, emarginate eye, securiform maxillary palp (MP), clypeus not distinct, apically blunt mandible. 2, Composite head in ventral view: left side showing protuberant eye, normal maxillary and labial palpi; right side showing non-protuberant eye, mandible with bifid apex, fringed prostheca (PR) and distinct molar part (MO). 3, Falcate mandible with retinaculum (R). 4, Filiform antennae. 5, Serrate ditto. 6, Pectinate ditto. 7, Flabellate ditto. 8, Clavate antenna with loose 3-segmented club. 9, Geniculate antenna with 1-segmented club.

C, cardo; CL, clypeus; GS, gular suture; L, labrum; LP, labial palp; M, mandible; MP, maxillary palp; MO, molar; MT, mentum; O, ocellus; P, pedicel; PM, prementum; PR, prostheca; R, retinaculum; S, scape; ST, stipes.

compound eves are sometimes more or less reduced or even absent, the corneal lenses of the ommatidia, forming their outer surface, may be very small and almost flat ("finely faceted eyes," more characteristic of sun-loving diurnal types) or larger and more or less strongly convex ("coarsely faceted eyes," generally found in nocturnal species); the outer surface of the eves may form a smooth continuation of the outline of the head (fig. 2, R. side), or they may be more or less markedly protuberant (fig. 2, L. side). The antennae have a fundamental number of 11 segments, often reduced to 10, 9, 8 or even less, but quite rarely increased to 12 or more, Segments 1 (scape) and 2 (pedicel) are usually more or less differentiated from the succeeding ones forming the flagellum; in geniculate antennae (fig. 9) the scape is very elongate and the rest of the antenna makes a sharp angle with it. segments of the flagellum are all more or less similar, elongate and cylindrical, the antenna is said to be filiform (fig. 4); if the flagellum resembles a string of beads it is called moniliform. If the segments of the flagellum are expanded on one side to give a more or less saw-like outline, the antennae are called serrate (fig. 5), if the outgrowths are longer they are said to be pectinate (fig. 6), if the outgrowths are extremely long the term is flabellate (fig. 7). Clavate (clubbed or capitate) antennae have from 1 to 5 of the apical segments enlarged and thickened and more or less sharply set off from the preceding ones (figs. 8, 9); the enlarged segments are normally dull and pubescent, bearing concentrations of sensillae. The antennal insertions vary from a completely lateral position under the sides of the frons (fig. 1, L. side) to a dorsal position on the front (fig. 1, R. Side)—in extreme cases the antennal insertions may only be separated by a narrow keel. The length of the antennae varies greatly, from much more than the whole length of the body to less than the width of the head. The mandibles are always distinct and generally of the opposed, biting type; they may be very slender and curved with sharp tips (falcate mandibles (fig. 3)), there may be a sharp tooth (retinaculum (fig. 3 R)) or a more or less soft hairy lobe (prostheca (fig. 2PR)) at the middle of their inner edge, the base of the inner edge may be expanded into a ridged or asperate crushing part (mola (fig. 2 mo)), the tips of the mandibles may be simple and acute, or blunt, or bifid, or bidentate, or The maxillae normally have all the typical parts (fig. 2), with a 4-segmented palp borne on a large palpiger, 2-segmented galea, and distinct lacinia; the apical segment of the palp may be enlarged and triangular (securiform palpi, fig. 1, R. side) with its distal margin wide and covered with a sensilla-rich membrane, or the apical segment may be very small and pointed (aciculate palpi, fig. 1, L. side); the galea may be palpiform, or hairy and lobiform, the lacinia is more usually pointed at the apex, or sometimes lacking (1-lobed maxillae). The labium is composed of two main parts, a distal one bearing the palpi and known as the prementum (mentum of some authors, fig. 2 PM), and a basal one articulating with the head capsule known as the mentum (submentum of some authors, fig. 2 MT): the labial palpi usually consist of two or three small segments, the terminal one of which may vary much as it does in the maxillary palpi; the prementum often bears in front a more or less membranous and bilobed ligula (fig. 2). Behind the labium lies the gular region of the head-capsule, bounded laterally by the gular sutures (fig. 2 gs) which are typically more or less parallel (fig. 2) but may converge or become partly or wholly confluent.

(2) The Prothorax (figs.10-13).—The term pronotum is usually applied only to the dorsal surface of the prothorax, which is commonly separated from the ventral surface by more or less sharp side-edges; the occurrence in Adephaga (and Archostemata) of noto-pleural sutures (fig. 10 NP) on the ventral surface some distance from the side-edges suggests that in Polyphaga too the pronotum should strictly be taken to include part of the hypomeron (ventral surface outside the sterno-pleural sutures, fig. 10 H). However, in this study the term pronotum will be used to refer to the dorsal surface only of the prothorax. The most consistent dividing lines on the ventral side of the prothorax are the sterno-pleural sutures (fig. 10 SP) which extend forwards from the outer angles of the front coxal cavities and separate



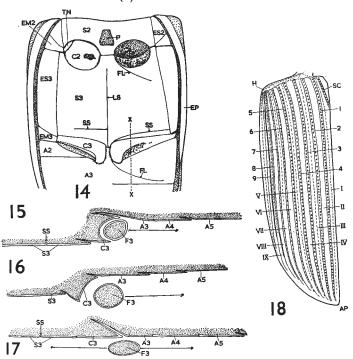
Fros. 10-13.—Prothoraces, ventral view. 10, Composite type, with ovate front coxae, left side (of figure) showing trochantin (TN) concealed, coxal cavity closed behind, no notopleural suture; right side with exposed trochantin, coxal cavity open (partially closed) behind, notopleural suture (NP) distinct. 11, Showing widely separated front coxae, the exposed parts of which are small and rounded but have hidden lateral extensions, cavities open behind, no distinct notopleural or sternopleural sutures. 12, Bostrychoid type with anterior orifice facing ventrally, front coxae (C1) projecting, their cavities partially closed behind, side edges (LK) distinct, sternopleural sutures obsolete. 13, Type with completely open front coxal cavities, strongly projecting front coxae with exposed trochantins, and distinct sternopleural sutures.

B, bar closing front coxal cavity internally; C1, front coxa; F1, front femora; H, hypomeron; IC, prosternal process; LK, side border of prothorax; NP, notopleural suture; PC, postcoxal process; S1, prosternite; SP, sternopleural suture; T, trochanter; TN, trochantin.

a median ventral sclerite—the prosternum (fig. 10 s1)—from the hypomera. The prosternum extends posteriorly between the front coxae as the prosternal process (fig. 10 IC), which may be of knife-edge thinness or even invisible externally in a few cases; its apex may extend little or not at all beyond the hind borders of the front coxae, or be prolonged and received into a pit in the mesosternum, or extend over and more or less conceal the latter sclerite. Processes of the hypomera (post-coxal processes, fig. 10 pc) may extend inwards behind the front coxae forming a partial (fig. 10, R. side) or, if they reach the prosternal process (fig. 10, L. side), a complete posterior closure of the front coxal cavities; these processes may incidentally more or less occlude the front pair of spiracles, which occur in the inter-segmental membrane in this vicinity. The front coxae may be of various sizes and shapes (figs. 10-13); often the outer part of an apparently small and rounded coxa is enclosed, so that it has a hidden lateral extension (fig. 11); the trochantins (fig. 10 tn) at the outer front angles of the coxae may or may not be externally visible.

(3) Mesothorax and Elytra (figs. 14-18).—The mesothorax in Coleoptera is almost always smaller than the prothorax and metathorax; when the insect is not in flight, the mesonotum is normally, except for a small median part known as the scutellum (fig. 31 sc), hidden by the elytra and the hind margin of the pronotum—the median anterior part of the mesonotum may sometimes bear a single or paired stridulatory files working against a ridge under the hind margin of the pronotum. The mesosternum (fig. 14 s2) is usually similarly shaped to the prosternum, with a median posterior intercoxal process and lateral wings in front of the middle coxae. The mesopleuron is composed of episternum (fig. 14 ES2) and epimeron (fig. 14 EM2), the former very rarely reaching the coxal cavities, the latter sometimes doing so (fig. 14, L. side) and sometimes not (fig. 14, R. side), in the latter case the middle coxal cavities are said to be closed outwardly (by the meeting of the mesoand metasterna). In some Adephaga the met-episterna (ES3) impinge on the middle coxal cavities, e.g. in DYTISCIDAE. The elvtra in beetles (fig. 18) frequently have 9 or 10 striae or rows of punctures with an additional short stria (scutellary striole, SC) close to the scutellum; the interstices and striae are usually numbered as shown in fig. 18—the alternate ones (I, III, V, VII, IX) are commonly underlain by tracheae and appear to represent primary wing-veins (respectively 2A, 1A, Cu, M, and R); they may bear articulated setae (e.g. in many CARABIDAE). The infolded costal margin of the elytron (also with a trachea, probably corresponding to C + Sc) is commonly delimited by a sharp outer edge, and is then known as the epipleuron (fig. 14 EP); the surface of the epipleuron may be flat and horizontal, or sloping downwards (inclined epipleura) or reflexed upwards and concave; it may extend the full length of the elytron or only part of it. The apices of the elytra in repose commonly form a conjoint rounded or slightly pointed outline, smoothly continuing the line of their outer edges, but they may be more or less sharply truncate at the apex, in which case they usually leave a part of the abdomen uncovered (cf. fig. 59). On the lower (inner) face of the elytron there may be a more or less strong flange some distance from the costal margin (e.g. in Dytiscidae and Curculionidae).

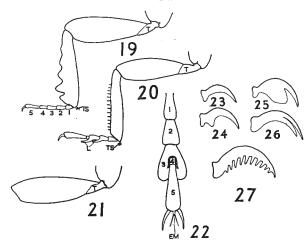
(4) THE METATHORAX AND WINGS.—In normal Coleoptera the metanotum is completely covered by the elytra in repose; in fully winged species



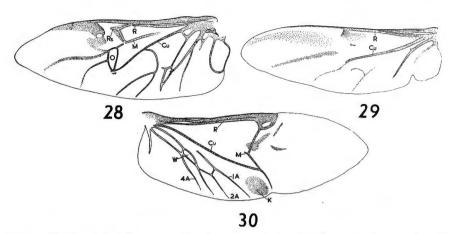
Figs 14-18.—Meso- and Metathorax. 14, Composite meso- and metathorax, ventral view: on left side mes-epimeron (EM2) reaching middle coxal cavity (so latter not closed outwardly by sterna), trochantin (TN) of middle coxae (C2) exposed. metasternum (S3) with transverse suture (SS) crossing middle line, met-episternum (ES3) parallel-sided, epimeron (EM3) exposed ventrally, hind coxae (C3) excavate posteriorly, extending laterally as far as outer edge of metasternum, dividing first visible abdominal sternite (A2), epipleuron (EP) extending whole length of elytron; on right side, middle coxal cavity closed outwardly by meeting of mesoand metasterna, metasternum with femoral line (coxal line, FL), its transverse suture (SS) not crossing middle line, metepisternum narrowed posteriorly, metepimeron not exposed ventrally, hind coxa not excavate posteriorly, extending laterally to meet elytral epipleuron, not dividing first visible abdominal sternite (A3), latter with femoral line (FL), epipleuron ending opposite first visible abdominal sternite. 15-17, Diagrammatic vertical longitudinal sections on a line corresponding approximately to XX in fig. 14; 15, showing excavate hind coxa (C3), first 2 visible abdominal sternites (A3-A4) connate, the third (A5) movably articulated; 16, showing somewhat projecting but not excavate hind coxa (C3), metasternum (S3) without transverse suture, first 3 visible abdominal sternites (A3, A4, A5) all movably articulated; 17, showing flat hind coxa, metasternum (S3) with transverse suture (SS), first 3 visible abdominal sternites (A3-A5) connate. 18, Typical elytron with punctured striae and scutellary striole, not truncate at apex, tracheae indicated by broken lines.

A2, A3, A4, A5, sternites of abdominal segments 2–5; AP, apical angle of elytron; C2, C3, middle and hind coxae; FL, femoral lines (coxal lines); EM2, EM3, meso- and meta-epimera; ES2, ES3, meso- and meta-episterna; EP, epipleuron; F3, hind femur; H, humeral protuberance; LS, longitudinal suture of metasternum; P, pit in mesosternum receiving tip of prosternal process; S2, S3, meso- and metasterna; SC, scutellary striole; SS, transverse suture of metasternum; TN, trochantin; X-X, line of section; 1–9, serial numbering of elytral striae; I-IX, serial numbering of elytral interstices. The arrows in 15–17 indicate range of movement of hind femur.

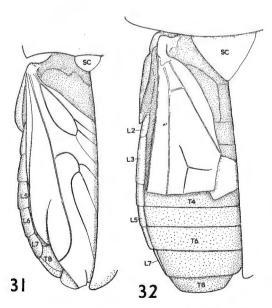
it is usually the largest of the thoracic terga, but undergoes reduction in flightless forms. The metasternum (fig. 14 s3) is normally a large, more or less rectangular sclerite, produced in the middle posteriorly as the inter-coxal process and emarginate on each side of its front edge to fit against the middle coxae. Its lateral margins may be parallel or more or less convergent anteriorly, and abut upon the elongate met-episterna (fig. 14 ES3) which in turn commonly meet the elytral epipleura in the resting position. met-epimera (fig. 14 EM3) are in large part hidden by the elytra, but portions of them may or may not be exposed at the outer ends of the hind coxae. The hind coxae (fig. 14 c3) are normally transverse and extend laterally at least as far as the outer edges of the metasternum, sometimes far enough to meet the elytral epipleura in repose (cf. L. and R. sides of fig. 14); their inner ends may be almost contiguous (i.e. inter-coxal process very narrow) or more or less widely separated (broad inter-coxal process). The apparent front margins of the hind coxae are usually fairly straight, less often convex. The main outer surface of the coxae may lie in one plane with the metasternum and first visible abdominal sternite (fig. 17)—giving "flat" hind coxae, or there may be an anterior horizontal part in the plane of the metasternum separated by an angle or ridge from an inclined posterior part sloping down (as seen from below) to the plane of the first visible abdominal sternite -" inclined" hind coxae (fig. 16), or the posterior part may become vertical and concave and separated from the horizontal part by a distinct flange— "excavate" hind coxae (fig. 15). The hind wings show three main types of venation (figs. 28, 29, 30); their folding patterns are characteristic in various



Figs. 19-27.—Legs. 19, Leg showing normal trochanter (T), dentate tibia, 5-segmented filiform tarsus. 20, Leg showing long trochanter (T), spinose tibia, pseudotetramerous tarsus with ventral lobes (L) on segments 2-3. 21, Base of leg showing heteromeroid trochanter. 22, Pseudotetramerous tarsus in dorsal view, showing small fourth segment connate to base of 5 and received in excavation of 3, large bisetose empodium (EM) between tarsal claws. 23, Simple claw. 24, Basally toothed claw. 25, Appendiculate claw. 26, Split claw. 27, Pectinate claw. EM, empodium: L, lobes of tarsal segments; T, trochanter; TS, tibial spur; 1-5, numboring of tarsal segments.



Figs. 28-30.—Wing Venation. 28, Adephagan type (Blethisa multipunctata). 29, Staphylinoid type (Pterolomaforsströmi). 30, Cantharoid type (Tetratomafungorum). Cu, cubitus; K, sub-cubital fleck; M, media; O, oblongum; R, radius; Rs, radial sector; W, Wedge cell (anal cell); 1A, 2A, 4A, 1st, 2nd and 4th anal veins.



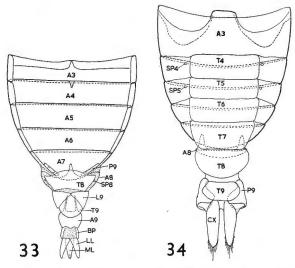
Figs. 31-32.—Wing folding patterns, left elytron removed. 31, Cantharoid type (Dascillus cervinus). 32, Staphylinoid type (Xylodrepa quadripunctata).

L2-L7, pleurites of abdominal segments 2-7; Sc, scutellum; T4, T6, T8, tergites of abdominal segments 4, 6, 8.

groups (cf. figs. 31, 32); in some species they may be reduced (brachyp-

terous) or even absent (apterous).

(5) THE LEGS.—The variation in the coxae has already been dealt with in treating the thoracic segments. The trochanters may vary in structure in different pairs of legs of the same beetle as well as between corresponding legs of different groups; their typical and presumably primitive form is illustrated in fig. 19—with the trochantero-femoral junction very oblique and the dorsal edge of the trochanter very short. In a number of groups we meet with "long" trochanters (e.g. Apion)) in which the trochantero-



Figs. 33-34.—Abdominal structure. 33, Male abdomen of a primitive Coleopteron, dorsal view with soft parts and tergites of segments 1-7 removed, genitalia extruded. 34, Female abdomen of a higher Coleopteron, soft parts and tergites of segments 1-3 removed, ovipositor extruded; coxites (CX) with only rudimentary styli at tips.

A3, A4, A5, A6, A7, A8, A9, sternites of abdominal segments 3 to 9; BP, basal piece of aedeagus; CX, coxite; LL, lateral lobe (paramere) of aedeagus; ML, median lobe (penis) of aedeagus; P9, paraproct (pleurite of ninth segment); SP4, SP5, SP8, spiracles of abdominal segments 4, 5, 8; T4, T5, T6, T7, T8, T9,

tergites of abdominal segments 4-9.

femoral junction is less oblique than usual (fig. 20) and the dorsal edge of the trochanter relatively long; in some forms the femur appears to extend dorsally round the trochanter to meet the coxae ("heteromeroid" trochanters, fig. 21). The femora may exhibit spines along the ventral edges, their shape is variable and the posterior pair are not infrequently inflated—commonly in connection with the development of the extensor tibialis muscle for jumping. The tibiae are usually broadened towards their apices, on the inner side of which they typically bear two (more rarely one or none) tibial spurs (fig. 19 TS); their outer edges may bear dentations (fig. 19) or articulated spines distinct from the general pubescence (spinose tibiae, fig.

20). The tarsi in Coleoptera provide many useful classificatory characters the most obvious of which is the "tarsal formula," which may be exemplified as x-y-z, where x represents the number of segments in the front tarsi, y that of the middle tarsi, and z that of the hind tarsi—the "y" is always the same as the x or the z, which themselves very rarely differ by more than 1. A tarsal formula of 5-5-5 would be described as pentamerous, 4-4-4 as tetramerous, 3-3-3 as trimerous; the term heteromerous could be taken to mean that any one pair of tarsi had a different number from the others, but is usually restricted to the formula 5-5-4. The tarsal formula sometimes differs in the sexes of the same species, in which case it is almost always the male which has the lower formula. Certain tarsal segments, particularly the basal and penultimate ones, are specially liable to reduction, and may become so small that they are only visible under high magnification; the intermediate (second to ante-penultimate) segments are liable to develop adhesive hairs below and to develop ventral extensions (lobes, see fig. 20L) which may themselves be bilobed apically (fig. 22). If segment 3 has such a lobe below while segment 4 is minute and connate to the base of 5, the tarsus is said to be pseudotetramerous (cryptopentamerous of some authors (fig. 22)); a similar tarsus with one fewer segment (hence formula 4-4-4 but appearing 3-3-3) is called pseudotrimerous. The tarsal claws may be simple (fig. 23). toothed (fig. 24), appendiculate (fig. 25), split (fig. 26), pectinate (fig. 27), unequal, connate basally, or reduced to one on each tarsus; there may be a bi- or plurisetose empodium (fig. 22EM) between the claws, or a pair of long lobiform (fig. 105) or bristle-like appendages below them.

#### THE COLLECTION AND IDENTIFICATION OF COLEOPTERA.

Adult beetles may be collected at all seasons of the year and in all types of habitat where any insect life is found. They may be killed in cyanide killing bottles such as are used by collectors of other insects, by exposure to the vapours of chloroform, ethyl acetate (acetic ether), and many other organic reagents, by brief immersion in boiling water, or by the old-fashioned laurel bottle. For the beginner, it is probably best to determine the specimens as far as the families as soon as they are killed, before mounting them in any way; this will be essential if it is desired to mount the specimens gummed flat on cards in the usual English style. If the specimens are pinned, or glued to the points of pieces of card in such a way that at least one side of the ventral surface can be seen, it should be possible to use the present key for determining the families at any later time. It is strongly advised that the beginner should acquire a few reliably identified examples of some of the commoner families as soon as possible, and check them against this key in order to familiarize himself with the characters used in it; if these suggestions are followed conscientiously at the beginning, it should not be long before the student finds himself able to dispense with the superfamily and family keys in the great majority of cases.

#### IMMATURE STAGES OF COLEOPTERA.

The larvae of beetles are almost as diverse in their habits and adaptations as the adults, and their study is a particularly interesting and rapidly developing branch of Coleopterology at the present time. There are many impor-

tant types of Coleoptera occurring in Britain whose larvae are quite unknown (e.g. Sphaerites, Sphaerius, Pseudopsis, Phloeophilus, Eugesthetus), many of our genera and the large majority of our species are in a similar position; the feeding habits of very many of our species are not or very inadequately Here then are fields in which amateur entomologists in Britain are in a position to do important original work. Keys to beetle larvae are not within the scope of the present work; students are recommended to refer to the papers by F. van Emden published in the Entomologists' Monthly The rearing of adult beetles from eggs or larvae is usually rather more difficult than the corresponding operation in Lepidoptera, and the techniques involved differ considerably in different groups of the order. Helpful information on this and other matters will be found in the Coleopterists' Handbook of the Amateur Entomologists' Society. Some species are much more easily found as larvae than as adults—in such cases rearing may be worth while from the ordinary collector's point of view, while to the serious entomologist the interest attaching to rearing experiments should be great enough to justify a considerable amount of effort in carrying them through.

#### GLOSSARY OF SPECIAL TAXONOMIC CHARACTERS USED.

Bostrychoid prothorax: characterised by a long and usually more or less humped notum, the anterior opening facing more downwards than forwards, the prosternum very short, e.g. in Anobiids, Ptinids, Cisids, Scolytids etc. (Figs. 12, 62.)

Clubbed antennae: with from 1 to 5 of apical segments broadened and marked off from the preceding ones by a distinct discontinuity, e.g. in Silphids and Crytophagids but not in *Byrrhus* or Bruchids which have the antennae simply thickened towards the apex. (Figs. 8, 9, 47, 74.)

Connate abdominal sternites: sternites between which the sutures have lost the usual telescoping mobility, e.g. first three sternites in Tenebrionids, first two in Curculionids, etc. (Figs. 15, 17.)

Dentate front tibiae: with the outer edges jagged or toothed, e.g. in Scarabaeoids, Histerids, most Scolytids, many Nitidulids, etc. (Fig. 19.)

Eighth abdominal segment: in Polyphagan beetles, where there are five visible abdominal sternites (ventrites) they belong to segments 3–7, where there are six ventrites the sternite of the eighth segment is also exposed. By lifting up the elytra of a relaxed specimen slightly it is usually easy to tell whether the last visible tergite belongs to the same segment as the last visible sternite or to the segment behind it. Chrysomelids normally have the eighth segment completely concealed (fig. 61), Hydrophilids and Elaterids will provide examples with only the tergite of the eighth segment exposed, Silphids and Staphylinids will illustrate exposed tergite and sternite of the eighth segment (fig. 60).

Excavate hind coxae: with a more or less vertical posterior face which is in some degree hollowed out to receive the retracted femur, e.g. in Dermestids or Elaterids. (Figs. 14, L. side, 15, 63, 65.)

Femoral plates: extensions of the postero-ventral edges of excavate hind coxac partially or wholly covering the retracted femora, e.g. in Byrrhids, Clambids, Trixagids, etc.

Femoral lines: raised ridges on the metasternum and first visible abdominal sternite extending backwards and outwards from inner ends of middle and hind coxae, as in Coccinellids, Biphyllids, etc. (Figs. 14, 91, 92.)

Flabellate antennae: with segments (except basal 2 and 3 and sometimes last) with long outgrowths on one side, usually at least half as long as whole

flagellum, e.g. in Rhipiphorids. (Figs. 7, 108.)

Flat hind coxae: main surface of hind coxa lying almost or quite in same plane as metasternum and first visible abdominal sternite, permitting a long unimpeded sweep of hind femora, e.g. in Dytiscids, Scarabaeids, Curculionids etc. (Figs. 17, 88, 92.)

Front coxal cavities completely open behind: propleuron (hypomeron) with no trace of processes projecting inwards behind front coxae, e.g. in

Elaterids, Cantharids. (Figs. 13, 63, 66.)

Front coxal cavities partially closed behind: propleuron with processes projecting inwards behind front coxae but not extending far enough to meet each other or the tip of the prosternal process, e.g. in many Heteromera (not Tenebrionids, Lagriids or Alleculids). (Figs. 11, 12.)

Heteromeroid trochanters: upper basal angle of femur extending round

Heteromeroid trochanters: upper basal angle of femur extending round dorsal side of trochanter and nearly or quite meeting coxa, e.g. in most

Heteromera. (Fig. 21.)

Long trochanters: femore-trochanteral junction not as oblique as usual, upper edge of trochanter almost or quite as long as width of trochanter at its junction with femur, e.g. in Apion, Cerylon, Anobiids, Ptinids. (Fig. 20.)

Pectinate antennae: segments (except from 2 to 4 of basal ones) with lateral outgrowths about as long as length of segments, e.g. in Pyrochroa,

Corymbites. (Figs. 6, 52, 107.)

Pseudotetramerous tarsi: appearing 4-segmented from extreme reduction of true fourth segment, segment 3 and usually 2 with ventral adhesive lobes at apex, e.g. in Chrysomelids, Curculionids. (Figs. 22, 109–111.)

Pseudotrimerous tarsi: similar to pseudotetramerous tarsi but with one

less basal segment, e.g. Coccinellids.

Securiform palpi: with last segment more or less hatchet-shaped or triangularly widened to its apex, e.g. in Coccinellids, many Heteromera etc. (Fig. 1, R. side.)

Serrate antennae: with segments (except basal 2 or 3) slightly expanded on one side only, giving a saw-like outline, e.g. in most Elaterids. (Fig. 5.)

Wing-venation: the Staphylinoid (figs. 29, 32) and Cantharoid (figs. 30, 31) types can usually be distinguished in the folded wing if an elytron is lifted up (figs. 31, 32); compare a Silphid with a Chrysomelid in this way. The same procedure permits observation of the terminal abdominal tergites.

#### THE CLASSIFICATION OF COLEOPTERA.

It would hardly be an exaggeration to say that it is impossible to find any two systematists working on Coleoptera who have used the same classification of the order. This state of affairs is confusing to the amateur and the non-specialist, and plays into the hands of those who accuse taxonomy of being a subjective and sterile pursuit of little or no scientific value. The system used in this part is true to this unfortunate tradition in that it does not follow those of any previous writers; I have published previously an

exposition of its theoretical basis (Crowson, 1955). For convenience of cross-reference, there is appended at the end of the part (p. 46) a conspectus of the system adopted, with indication of the names of the corresponding groups in Fowler's Coleoptera of the British Islands, Reitter's Fauna Germanica: Käfer, Joy's Practical Handbook of British Beetles, Imm's General Textbook of Entomology, and Kloet and Hincks' Check List of British Insects (the Coleoptera section of which does not itself follow any previously published system).

#### THE SUBORDERS OF COLEOPTERA.

The suborders Adephaga and Polyphaga have been recognised by nearly all serious students of classification in beetles during the present century; the addition of a third (Archostemata, not represented in Europe) has become fairly widely accepted in recent years. As first recognised by Forbes (1926) there are some grounds for separating three small families (Cyathoceridae, Sphaeriidae and Hydroscaphidae) from Polyphaga to form a fourth suborder, which I have elsewhere named Myxophaga (Crowson 1955). In the present work, however, these forms will be treated as a superfamily in Polyphaga. The two generally recognised suborders in Britain may be distinguished thus:

#### Suborder 1.—ADEPHAGA.

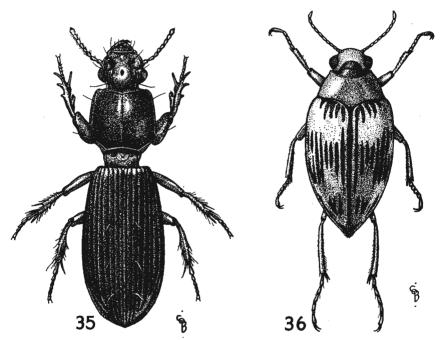
A single superfamily Caraboidea.

#### CARABOIDEA.

- Antennal insertions somewhat approximated on frons, clypeus extending laterally in front of them (fig. 37); both spurs of front tibiae quite terminal, no trace of an antenna-cleaning excavation on front tibia; side margins of prothorax obsolete; length at least 9 mm..............................Subfamily 1. CICINDELINAE
- Antennal insertions not so approximated, clypeus not extending in front of them..3

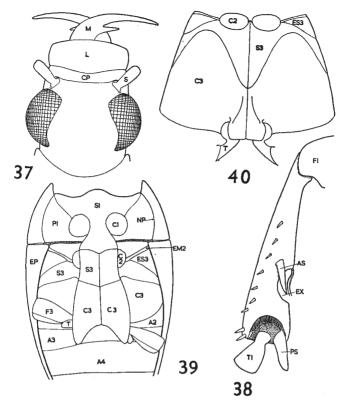
  Antennae with segments 1-4 glabrous; metepimeron normally not exposed ventrally; antenna-cleaning excavation on inner side of front tibia often not or little developed; front coxal cavities often open behind
  - Subfamily 2. Carabinae Antennae with not more than three basal segments glabrous; metepimeron always exposed ventrally (fig. 14, L. side); front tibiae always with well-developed antenna-cleaning excavation (fig. 38); front coxal cavities closed behind.....4

4. Mesepimera reaching middle coxal cavities (fig. 14, L. side); species of fossorial adaptations with pedunculate mesothorax (fig. 35) and strongly dentate front tibiae; antennae rather stout and inserted in deep grooves



Figs. 35-36.—35, Clivina fossor L.; 36 Haliplus obliquus F.

Postero-ventral edgs of hind coxae produced into large plates covering basal abdominal sternites, elytra with large punctures in regular rows, facies charac-Postero-ventral edges of hind coxae not so developed; elytra without such regular Hind coxae relatively small, metasternum with a distinct transverse suture in front of them (cf. fig. 14, L. side); head strongly exserted with very convex eyes; species with very convex dorsal and ventral surfaces and fairly large size (length 8-10 Hind coxae much larger (figs. 39-40), metasternum without a transverse suture; head less exserted, eyes not or little projecting beyond its general outline.....8 Upper surface very convex, lower surface almost flat; antennae short, somewhat thickened; scutellum hidden; metasternum and hind coxae characteristically shaped (fig. 39)......4. Noteridae If upper surface so convex, lower surface also very convex; antennae not thickened, usually longer; metasternum and hind coxae differently shaped (fig. 40) 5. DYTISCIDAE



Figs. 37-40.—37, Cicindela campestris L., antero-dorsal view of head. 38, Harpalus sp., R. front tibia, anterior view. 39, Noterus clavicornis Deg., thorax, ventral

view. 40, Laccophilus minutus L., metathorax, ventral view.

A2, A3, A4, visible abdominal sternites 1-3; AS, anterior spur; C1, C2, C3, front, middle and hind coxae; CP, clypeus; EM2, mes-epimeron; EP, elytral epipleuron; ES3, met-episternum; EX, antenna cleaning excavation; F1. F3. front and hind femora; L, labrum; M, mandible; NP, notopleural suture; PS, posterior spur; S, scape of antenna; S1, S3, pro- and metasterna; T1, basal tarsal segment; T, trochanter.

#### Suborder 2.—POLYPHAGA.

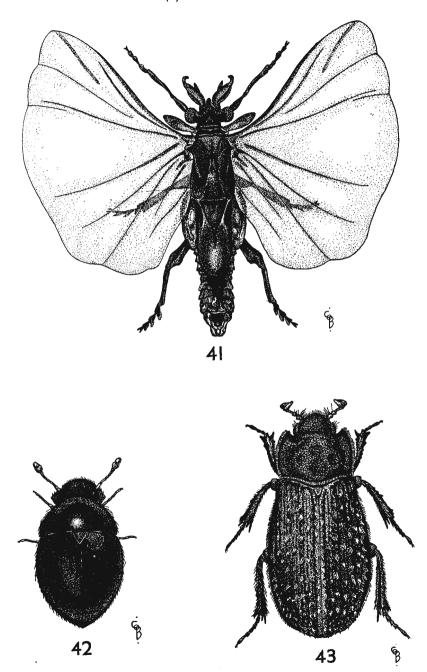
#### KEY TO SUPERFAMILIES.

Male: elytra reduced to small haltere-like structures (fig. 41), wings fully developed prothorax small, head completely exserted; antennae 5-7 segmented, at least segment 3 with a flattened lateral outgrowth; tarsal formula 4-4-4 or 3-3-3, claws absent. Female: apterous and legless, remaining in puparium in body of larval host. (Order Strepsiptera auctt.)..................19. Stylopoidea (p. 45) Mule with elytra not or less reduced, prothorax larger; female always with legs....2

Abdomen with only three visible sternites; prothorax with distinct noto-pleural sutures; hind coxae with very large femoral plates; antennae clubbed; species

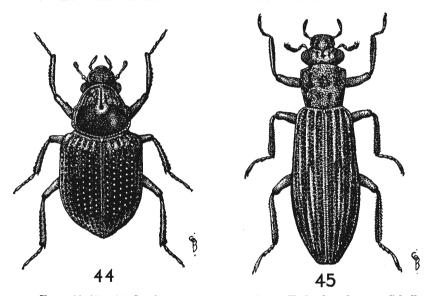
very small (length less than 1 mm.), of very convex short oval form

1. Sphaerioidea (p. 26) Abdomen with more than three visible sternites; prothorax without distinct noto-



Figs. 41-43.—41, Stylops sp., male. 42, Clambus pubescens Redt. 43, Trox scaber L.

- 3. Head very large, nearly or quite as broad as prothorax, capable of being reflexed against underside of thorax; hind coxae with very broad femoral plates; antennae 10-segmented with 2-segmented club; tarsal formula 4-4-4, legs slender; species of short ovate and convex form (fig. 42)
  - 6. Dascilloidea in part (p. 30)
    If head nearly or quite as wide as prothorax, hind coxae without femoral plates
    and antennae different......4
- 4 Antennae usually 9-10 (rarely 11) segmented, the apical 3-5 (rarely 6 or 7) segments expanded on one side to form a lamellate club (fig. 46), segment 8 not smaller than 7; tarsal formula almost always 5-5-5, segments never lobed below; front tibiae with outer edge more or less dentate (cf. fig. 19), only one apical spur; front coxae very large, transverse or projecting, hind coxae very large, quite flat (cf. fig. 17); general form stout, length at least 2.5 mm.

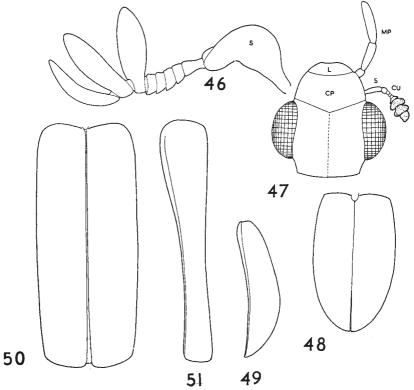


Figs. 44-45.—44, Georissus crenulatus Rossi. 45, Hydrochus elongatus Schall.

5 Antennae very short, usually less conspicuous than the prominent maxillary palpi (fig. 47), 7-9 segmented, the last 3-5 segments forming a dull pubescent club; tarsi filiform, formula 5-5-5 but sometimes appearing 4-4-4 through extreme reduction of basal segment; tibiae more or less spinose on outer edges (cf. fig. 20); species usually glabrous and strongly convex above, under side more or less flat and with characteristic close tomentum; clypeus large, usually extending over antennal insertions (fig. 47), vertex usually with median impressed line

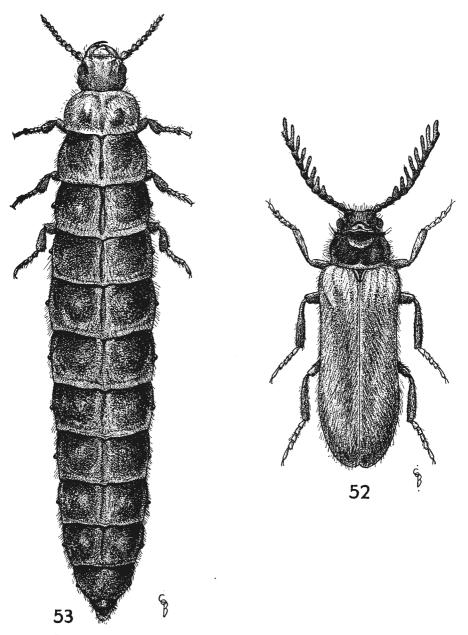
2. Hydrophiloidea (p. 26)

Antennae more or less clubbed, often geniculate, inserted in pits or grooves; head more or less rostrate, gular sutures nearly always obsolete or confluent (fig. 110); middle coxal cavities almost always closed outwardly by sterna; abdomen with at least two basal sternites more or less connate....18 Curculionoidea (p. 43)



Figs. 46-51.—46, Scarabaeid antenna. 47, Hydrophilid head, dorsal view. 48, Helodid elytra, dorsal view. 49, Ditto, lateral view. 50, Cantharid elytra, dorsal view. 51, Ditto, lateral view.

CP, clypeus; CU, cupule; L, labrum; MP, maxillary palp; S, scape.

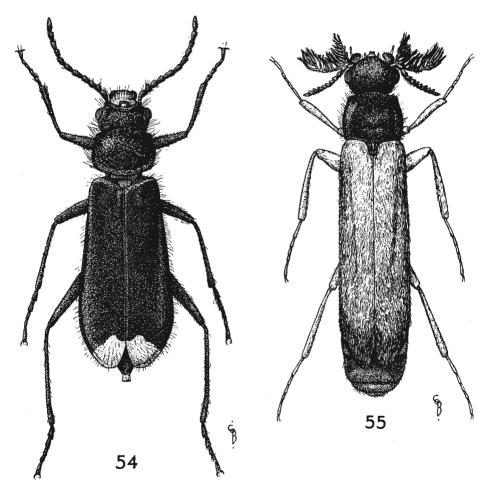


Figs. 52-53.—52, Drilus flavescens Geoff., male. 53, ditto, female.

9 Projecting bristly hairs present on some parts of the body, usually on margins of prothorax and elytra (fig. 54); coloration often partly metallic, elytra often spotted or banded; front coxal cavities often partially closed behind; both sexes fully winged, elytra always covering folded wings. 14. Cleroidea (p. 34) Projecting bristly hairs absent; coloration very rarely metallic, elytra rarely spotted or banded.......10 10. Tarsi very long and thin, filiform; antennae short, serrate (fig. 55); females fully winged, males with extraordinarily developed maxillary palpi (fig. 55);

general form cylindrical, hardly at all flattened.....15. Lymexyloidea (p. 34)

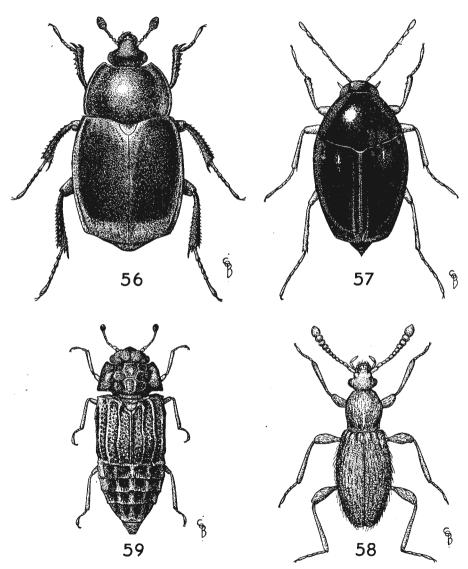
Tarsi shorter, penultimate segment more or less lobed below; antennae longer; 



Figs. 54-55.—54, Malachius bipustulatus L. 55, Hylecoetus dermestoides, L., male.

Apices of elytra forming a smooth slightly acuminate outline (fig. 48), fairly strongly and smoothly deflexed in side view (fig. 49); general form broader and more convex dorsally; hind coxa distinctly excavate; five visible abdominal sternites
 Dascilloidea (p. 30)

Apices of elytra more or less truncate or obtuse (fig. 50), not smoothly deflexed in side view (fig. 51); general form narrower, less convex; hind coxae not distinctly excavate; usually six to seven visible sternites.....11. Cantharoidea (p. 32)



Figs. 56-59.—56, Sphaerites glabratus F. 57, Scaphisoma agaricinum L. 58, Neuraphes elongatulus Muell. 59, Micropeplus porcatus F.

Tarsi 5-5-4 segmented in both sexes; antennae never geniculate, nor with Anisotomid type of club (cf. fig. 75); elytra, if truncate, never leaving more than one abdominal tergite uncovered (Heteromera auett.)......16. Cucujoides (p. 34) If tarsal formula 5-5-4 in both sexes, elytra truncate leaving two abdominal tergites uncovered, or antennae with Anisotomid type of club (fig. 75)......13 Elytra truncate leaving at least one, usually two, abdominal targitos uncovered; species with hard integument, glabrous above, usually black and shining (fig. 56); tarsi without lobed segments; front tibing dentate (fig. 19) or spinose outwardly; hind coxae flat; abdomen with five visible sternites, the last visible tergite belonging to same (seventh) segment as last visible sternite (cf. fig. 61) 3. Historoldea (p. 27) If elytra truncate leaving one or more tergites uncovered, tarsi with lobed segments or abdomen with at least six visible sternites......14 14 Elytra truncate leaving at least three abdominal tergitor not covered; antennae rarely with apical segments forming a well-marked club; tarsi usually without lobed segments; tergite and sternite of abdominal segment 8 (corresponding to sixth or seventh visible sternite) exposed; front coxue more or less projecting 4. Staphyllnoidea in part (p. 27)

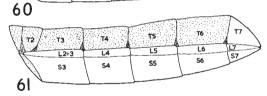
T2. T3 T4 T5 T6 T7 T8

L3 L4 L5 S9

S3 S4 S5 S6 S7

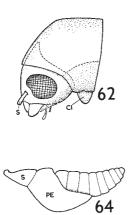
L7

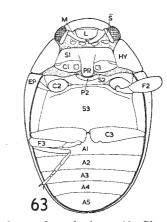
If elytra truncate leaving three or more tergites uncovered, antennae strongly



Figs. 60-61.—Abdomens, lateral view. 60, Haplogastran abdomen, showing separate pleurite (L2) of segment 2, and exposed tergite and sternite (T8, S8) of segment 8; spiracles of segments 2—8 indicated. 61, Symphiogastran abdomen, showing no separate pleurite of segment 2, tergite and sternite of segment 8 concealed, spiracles of segments 2-7 indicated.

L, pleurites; S, sternites; T, tergites—numbered according to segments.

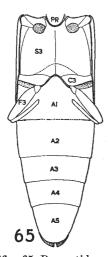


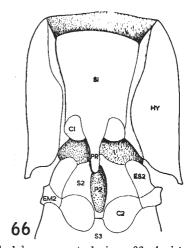


Figs. 62—64.—62, Bostrychoid head and prothorax, lateral view. 63, Simplocaria semistriata F., whole insect, ventral view (most of legs removed). 64, Dryops auriculatus Geoffi., antenna.
A1, A2, A3, A4, A5, visible abdominal sternites (ventrites) 1-5; C1, C2, C3, promeso- and meta-thoracic coxae: F2, F3, middle and hind femora: EP, elytral

A1, A2, A3, A4, A5, visible addominat sterines (ventries) 1-3; C1, C2, C3, piot, meso- and meta-thoracic coxae; F2, F3, middle and hind femora; EP, elytral epipleuron; HY, hypomeron; L, labrum; M, mandible; P2, pit in mesosternum receiving tip of prosternal process; PE, pedicel; PR, prosternal inter-coxal process; S, scape; S1, S2, S3, pro-, meso- and meta-sterna.

Prosternal process not deeply received in monomeroum, middle coxac not widely separated; antennae strongly clubbed; turni without lobed segments; head often with median occllus; elytra, if striate, with a soutellary stricte



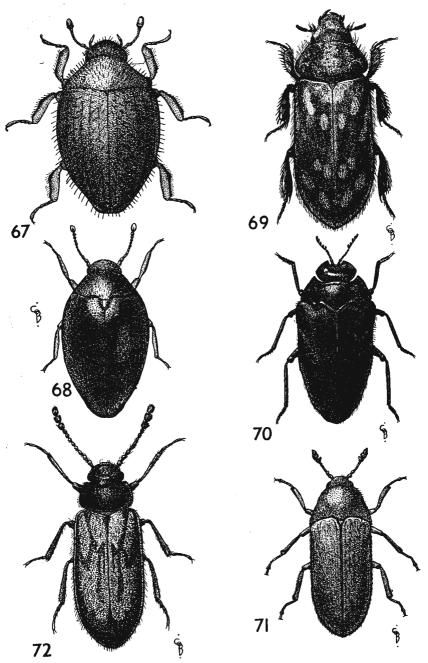


Figs. 65-66.—65, Buprestid metathorax and abdomen, ventral view. 66, Agrictes sp. pro- and mesothorax, ventral view.

A1, A2, A3, A4, A5, visible abdominal sternites 1-5; C1, C2, C3, front, middle

A1, A2, A3, A4, A5, visible abdominal sternites 1-5; C1, C2, C3, front, middle and hind coxae; EM2, mes-epimeron; ES2, mes-episternum; F3, hind femur; HY, hypomeron; P2, pit in mesosternum receiving tip of prosternal process; PR, prosternal inter-coxal process; S1, S2, S3, pro-, meso- and meta-sterna.

25 Front coxae projecting; at least some part of body with outstanding bristly hairs (cf. fig. 54); species never black; elytra completely covering abdomen



Figs. 67–72.—67, Syncalypta striatopunctata Steff. 68, Limnichus pygmaeus Sturm. 69, Heterocerus obsoletus Curt. 70, Trachys minuta L. 71, Trixagus dermestoides L. 72, Phloiophilus edwardsi Steph.

#### Superfamily 1.—SPHAERIOIDEA.

I have elsewhere (Crowson 1955) treated this group as constituting a suborder (Myxophaga mihi) of its own; of the three families constituting it, only one is represented in Britain. Another British genus (Calyptomerus Redt.) has very similar wing-folding and venation to that of Sphaerioidea, and on that account I have previously (loc. cit.) assigned it to this superfamily, but so far no definite confirmation of this has been forthcoming from other structural features of the imagines or of the larvae (which have recently been bred); for the present it seems best to leave Calyptomerus near Clambus.

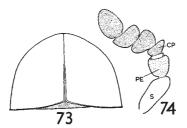
#### Family 1.—SPHAERIIDAE.

The family comprises a single genus *Sphaerius* Waltl, represented in Britain by *S. acaroides* Waltl. The species has been found under pieces of wood, etc. in wet places on areas of alkaline peat (the true Fons of the plant ecologists) in Central and Eastern England, but I have been able to find no records of its occurrence during the last thirty years.

#### Superfamily 2.—HYDROPHILOIDEA.

Certain Anisotomidae (Staphylinoidea) might be mis-traced here; they never exhibit the characteristic large clypeus of the Hydrophiloidea (fig. 47) and their antennal club is usually characteristic (fig. 75).

1 Antennal club 5-segmented; abdomen with six or seven visible sternites; size small (length less than 3 mm.); species not of very convex or rounded from



Figs. 73-74.—73, Spercheus emarginatus Schall., outline of elytra viewed from behind. 74, ditto, antenna. S, scape; PE, pedicel; CP, cupule.

Front coxae very large and projecting, concealing prosternum; tarsi 4-4-4 segmented; first two visible abdominal sternites connate; antennal insertions not hidden by sides of clypeus; species small, of characteristic shape (fig. 44)
 Georissidae

5. Hydrophilidae

#### Superfamily 3.—HISTEROIDEA.

Certain NITIDULIDAE (Cucujoidea) might be referred here; they may be distinguished by having tarsal segment 3 always somewhat lobed and with a pubescent pad below. PLATYPODIDAE (Curculionoidea) have much larger and more projecting front coxae and a more deflexed head (fig. 115).

abdominal sternites uncovered; antennae geniculate, club often solid (fig. 9); front tibiae dentate outwardly (cf. fig. 19)................2. HISTERIDAE

#### Superfamily 4.—STAPHYLINOIDEA.

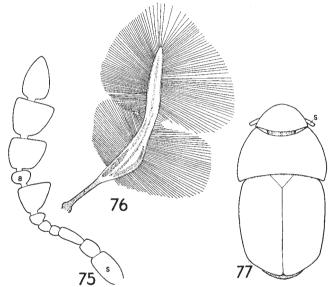
If the wings are not examined, certain Cucujoidea (e.g. some Myceto-PHAGIDAE) might be mistraced here; some NITIDULIDAE may be distinguished by their lobed third tarsal segment and strongly clubbed antennae; certain Cantharoidea with reduced elytra and some MELYRIDAE (Cleroidea) might be sought here if their Malacoderm features have been overlooked. THORIC-TIDAE (Dermestoidea—fig. 85), if traced to this superfamily, would be taken to the vicinity of SCYDMAENIDAE, from which Thorictodes differs inter alia in having a very broad and compact 3-segmented antennal club. Clambidae (Dascilloidea, fig. 42) and SPHAERIIDAE (Sphaerioidea) were formerly referred to Staphylinoidea but both differ in having very broad femoral plates to the hind coxae. The Cucujoid CORYLOPHIDAE, also commonly grouped here hitherto, are included in the key below; MICROPEPLIDAE are treated as a family here but were treated as a subfamily of STAPHYLINIDAE by Tottenham in the present series. If the elytra can be lifted up, Staphylinoid beetles may usually be recognised by the Staphylinoid wing-venation and folding (figs. 29, 32), and by the Haplogastran type of abdomen, with the pleurites of abdominal segments 2 and 3 separate (figs. 32, 60, cf. 31, 61). This type of abdominal structure is also found in the two previous superfamilies, and in the Scarabaeoidea. All these groups have on this account been placed in series Haplogastra by some.

1 Species very small (length rarely more than 1 mm.), prothorax large and semicircular in outline (fig. 95), partly or wholly hiding head; hind coxae widely separated, without trace of femoral plates; elytra somewhat truncate but almost or quite covering abdomen; tarsal formula 4-4-4, more or less pseudotrimerous (Superfamily Cucujoidea fam. CORYLOPHIDAE)

2 Species very small (length not more than 1.3 mm.), antennae slender with segments 1-2 thick and a loose 3-segmented club, segments bearing whorls of projecting hairs; hind coxae with femoral plates on inner half; all coxae more or less widely separated; wings characteristic (fig. 76), fringed with long hairs

Eyes greatly reduced; head with a narrow neck; species of broad rather parallel-sided and flattened form (fig. 77); front coxae small, reunded; turnal formula 5-5-5; antennae with segments 7-11 slightly enlarged, 8 smaller than 7 and 9

(Leptinus Mueller, L. testuceus Muell.)

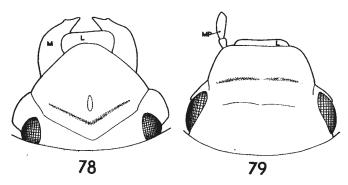


Figs. 75-77.—75, Anisotomid antenna. 76, Ptillid wing. 77, Leptinus testaceus, outline of body. S, scape. 8, eighth segment of antenna.

- 8 General shape characteristic (fig. 57), very convex dorsally and ventrally; species glabrous and very shining; antennae with weak 5-segmented club
  - Shape different; elytra normally leaving more than one abdominal tergite uncovered; antennae usually filiform......9
- 9 Antennae 9-segmented with 1-segmented club; prothorax and elytra with characteristic raised ridges (fig. 59); front coxae transverse, scarcely projecting
  - 7. MICROPEPLIDAE
    Antennae nearly always 11-segmented, usually filiform; front coxae more or less
    projecting; prothorax and elytra without such raised ridges......10
- 10 Abdomen fully flexible; tarsal formula rarely 3-3-3, claws almost always equal; body rarely with deep cuticular pits; general form almost always more elongate
  8. Staphyllnidae

#### Superfamily 5.—SCARABAEOIDEA.

Certain Anisotomidae and perhaps some Hydrophilidae-Sphaeridiinae might be referred here; neither group has the dentate front tibiae of Scarabaeoids; the characteristic antennal club of the Anistomids (fig. 75) and the head structure (fig. 47) of the Hydrophiloids will usually suffice to distinguish them.



Figs. 78-79.—78, Geotrupes vernalis L., head, antero-dorsal view. 79, Amphimallon solstitialis L., head, antero-dorsal view. L., labrum; M, mandible; MP, maxillary palp.

#### Superfamily 6.—DASCILLOIDEA.

CLAMBIDAE are transferred here from Staphylinoidea on account of the evident affinity shown by adult and larval *Clambus* to the non-British genus *Eucinetus*, which itself has affinities to Helodidae. In the present state of uncertainty over the true relationships of *Calyptomerus*, it seems best to leave the genus in Clambidae. *Eubria*, included under Helodidae in almost all previous British works, is here transferred to the Dryopoid family PSEPHENIDAE.

#### Superfamily 7.—BYRRHOIDEA.

#### With a single family, BYRRHIDAE.

Most previous authors have included here the genus Limnichus (see fig. 68) (Dryopoidea: Limnichide), which, as pointed out by Hinton (1938), may be separated from Byrrhided by the distinct fronto-clypeal suture inter alia. The Sphindid genus Aspidiphorus (also placed here by Fowler) is at once separable from Byrrhids by the partially closed front coxal cavities and the completely un-excavate hind coxae. The Buprestid Trachys (fig. 70) has strongly lobed tarsal segments and serrate antennae. Some similarly shaped Cucujoids (e.g. Ephistemus), Chrysomeloids (e.g. Lamprosoma) and Hydrophiloids (e.g. Megasternum), etc., may be excluded from Byrrhoidea by the flat hind coxae.

#### Superfamily 8.—DRYOPOIDEA.

- 4. Form very convex, Byrrhid-like (fig. 68); hind coxae excavate; upper surface finely pubescent; last tarsal segment shorter than preceding ones taken together

  3. LIMNICHIDAE

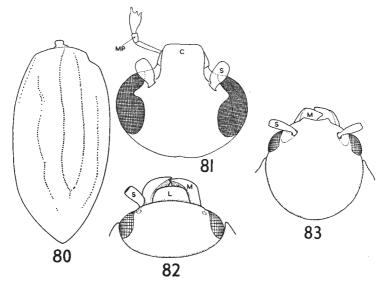
(Limnichus Latr., L. pygameus Sturm)

#### Superfamily 9.—BUPRESTOIDEA.

With a single family, BUPRESTIDAE.

#### Superfamily 10.—ELATEROIDEA.

This superfamily and the last together form the old group Sternoxia.



Figs. 80–83.—80, Eubria palustris Germ., L, elytron, dorso-lateral view. 81, Eubria palustris Germ., head, antero-dorsal view. 82, Cryptohypnus riparius F., head, antero-dorsal view. 83, Melasis buprestoides L., head, antero-dorsal view. C, clypeus; L, labrum; M, mandible; MP, maxillary palp; S, scape.

#### Superfamily 11.—CANTHAROIDEA.

The Dermestid genus *Thylodrias* might be traced here; it differs from all Cantharoids in having a median dorsal occllus and some differentiation of the apical antennal segments. The Cleroid Melyridae (which greatly resemble the present group in the external features of the adults, and have generally been placed in it hitherto) can be distinguished either by the presence of long appendages below the tarsal claws (cf. fig. 105) or by having outstanding bristly hairs on some parts of the body (cf. fig. 54).

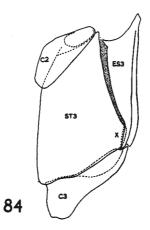


Fig. 84.—Silis ruficollis F., metasternum, latero-ventral view. C2, C3, middle and hind coxae; ES3, met-episternum; ST3, metasternum; X, angulation of side border of metasternum.

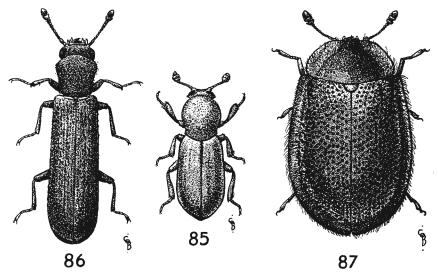
Luminous organs usually present on posterior abdominal segments; sexual dimorphism strong, females without wings and with elytra reduced or absent, males with very large eyes partially concealed under front margin of pronotum; side margins of metasternum straight; trochanters normal....2. LAMPYRIDAE Luminous organs absent; sexual dimorphism slight or absent; side margins of metasternum characteristically bent (fig. 84) or trochanters long (cf. fig. 20)....3

4. LYCIDAE

#### Superfamily 12.—DERMESTOIDEA.

1 Hind coxae small, not appearing to extend as far laterally as margins of metasternum; hind angles of prothorax blunt (fig. 85); ocellus absent; ventral surface without silky pubescence; size small (length less than 1.5 mm.)

2. THORICTIDAE (Thorictodes Reitt., T. heydeni Reitt.)



Figs. 85-87.—85, Thorictodes heydeni Reitt. 86, Lyctus brunneus Steph. 87, Thymalus limbatus F.

2 Antennae various, never with 3-segmented serrate club; head with median dorsal ocellus, or lower surface with close silky pubescence; prosternal process not or slightly received in mesosternum; elytra, if striate, with a scutellary striole...2

1. Dermestidae

Antennae with 3-segmented serrate club; dorsal ocellus absent, lower surface without such silky pubescence; prosternal process broadly and deeply received in mesosternum; shape characteristic (fig. 71)...(Elateroidea fam. TRIXAGIDAE)

#### Superfamily 13.—BOSTRYCHOIDEA.

This grouping, often together with Lymexyloidea and CISIDAE, comprised the Teredilia of older authors. The Hydrophiloid *Georissus* might be mis-traced here; its facies (fig. 44) and water-side habitat should suffice to distinguish it. Many Scolytidae (Curculionoidea) much resemble Bostrychidae; they may usually be separated by their tarsi with segment 4 (instead of 1) greatly reduced and by their completely closed front coxal cavities. The Cucujoid CISIDAE and SPHINDIDAE differ from Bostrychids notably in having a strongly bordered prothorax.

3 Front coxee not at all projecting, their cavities completely closed behind; antennae 11-segmented with 2-segmented club; base of prothorax (fig. 86) much narrower than elytra at shoulders................................4. LYCTIDAE Front coxee more or less projecting (fig. 12); antennae never with 2-segmented

Front coxae more or less projecting (fig. 12); antennae never with 2-segmented club; base of prothorax not or scarcely narrower than elytra at shoulders

3. Bostrychidae

### Superfamily 14.—CLEROIDEA.

## Superfamily 15.—LYMEXYLOIDEA.

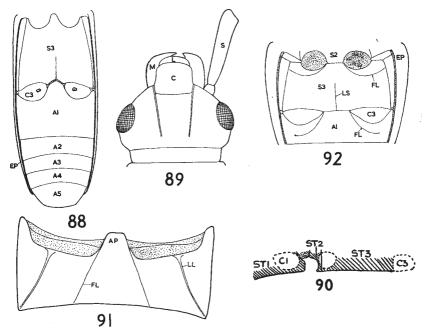
The single family LYMEXYLIDAE (LYMEXYLONIDAE auctt.) has generally been classed with either "Malacodermata" or "Teredilia," but shows much closer relationships to Cucujoidea. Some Melyridae (Cleroidea) might be referred here, but may be distinguished by the appendages below the tarsal claws and by the different type of colouration (brownish to fuscous in Lymexylids).

## Superfamily 16.—CUCUJOIDEA.

A very large and diverse group, comprising the Heteromera plus most of the Clavicornia of older authors. A number of genera belonging to other superfamilies might be mis-traced here, e.g. the Anisotomid *Triarthron* which has an ordinary 3-segmented antennal club. This genus differs from any Cucujoidea in combining very transverse front coxae, filiform 5–5–5 tarsi, and spinose front tibiae. Another Staphylinoid, *Leptinus*, may be recognised by its general form (fig. 77) and by combining filiform 5–5–5 tarsi with vestigial eyes. The Bostrychoid *Lyctus* also has a distinctive facies (fig. 86); the combination of completely filiform 5–5–5 tarsi with segment 1

minute, a 2-segmented antennal club, the prothorax markedly produced over the head, and middle coxal cavities closed outwardly by the sterna, will separate it from any Cucujoidea. The Cleroid Trogositidae are very Clavicorn-like (e.g. Thymalus, fig. 87); the completely filiform 5–5–5 tarsi with segment 1 very small and a large bisetose empodium between the claws, together with the very transverse front coxae and the 9–10 segmented antennae with an asymmetrical club, should suffice for their recognition. Phloiophilus (fig. 72), here attributed to Cleroidea, has also possible affinities to the Cucujoid Biphyllidae and Byturidae.

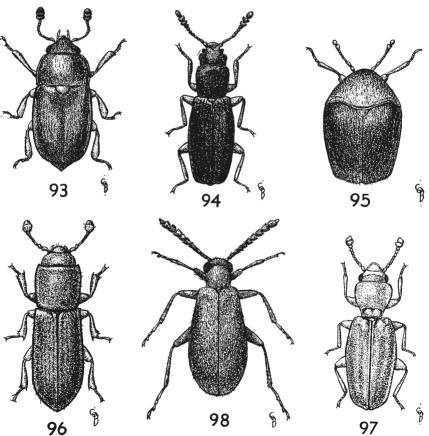
Tarsal formula 5-5-4 in both sexes; antennal club often weak or absent; tarsi often with penultimate segment more or less lobed, rarely of pseudotetramerous type; prothorax often without distinct side borders (Heteromera auctt.).....21



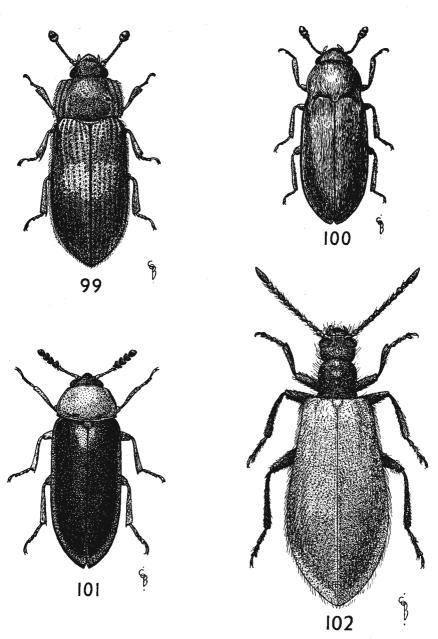
Figs. 88–92.—88, Rhizophagus bipustulatus F., metathorax and abdomen, ventral view 89, Pediacus dermestoides F., head, dorsal view. 90, Phalacrus sp., diagrammatic median longitudinal section of thoracic venter. 91, Biphyllus lunatus F., 1st abdominal sternite, ventral view. 92, Coccinellid metathorax and base of abdomen, ventral view. A1, A2, A3, A4, A5, visible abdominal sternites 1–5; AP, anterior inter-coxal process of basal abdominal sternite; C, clypeus; C1, C2, C3, front, middle and hind coxae; EP, elytral epipleuron; ES2, mes-episternum; FL, femoral lines; L, labrum; LL, lateral line; LS, longitudinal suture of metasternum; M, mandible; PC, post-coxal process of hypomeron; S, scape; S1, S2, S3, pro-, mesoand meta-sterna.

2	Elytra truncate leaving last abdominal tergite uncovered; antennae 10-segmented with 1-segmented club; form rather elongate and parallel-sided; first visible abdominal sternite considerably longer than following ones (fig. 88); tarsal formula 5-5-4 in males, ante-penultimate segment very slightly lobed below  2. RHIZOPHAGIDAE
	If elytra truncate leaving last abdominal tergite uncovered, antennae and tarsi different
3	Front coxae very transverse with large exposed trochantins (cf. fig. 10 r side), middle coxae also with large trochantins; antennae with strong 3-segmented club; elytra often more or less truncate (fig. 93); tarsal segments 2-3 more or less lobed below, formula 5-5-5 in both sexes; outer edges of front tibiae more or less dentate
4	Tarsal formula 5-5-5 (sometimes 5-5-4 in males), penultimate segment often more or less reduced and ante-penultimate lobed below
5	Tarsal formula 4-4-4 (sometimes pseudctrimerous) or 3-3-3
	short, anterior face of mesosternum concave to receive the ventro-apical portion of the prosternum (fig. 90); antennae with compact 3-segmented club; tarsi pseudotetramerous
6	Pronotum with characteristic raised longitudinal lines at sides (fig. 99); antennae clubbed; front coxae transverse, their cavities visibly closed behind; trochanters
	heteromeroid (fig. 21); first visible abdominal sternite with femoral lines; elytral epipleura well-marked
	elytral epipleura well-marked
7	Front coxae transverse with exposed trochantins, their cavities visibly closed behind; trochanters heteromeroid (fig. 21); upper surface closely pubescent (fig. 100); elytra without distinct striae, epipleurae distinct at least in basal
	half; tarsi pseudotetramerous
8	Antennes 10 as green to describe a result of 2 as green to desk a torrise open bening
o	Antennae 10-segmented with weak 2-3-segmented club; tarsi without lobed segments; front coxae transverse; form not flattened; elytra with indistinct epipleurae, scutellary striole distinct6. Sphindidae
	Antennae nearly always 11-segmented; if front coxae transverse and elytra without distinct epipleurae, elytra without distinct scutellary striole9
9	Front coxae small, rounded, their cavities broadly closed behind: side edges of prothorax more or less dentate; general form elongate and more or less depressed; antennae rather short and thick, club weak or absent; tarsal formula 5-5-5 in both sexes, segment 4 smallest, 3 more or less lobed below5. SILVANIDAE
	If front coxae small and rounded and with closed cavities, sides of prothorax not dentate
10	Tarsi with segment 1 much shorter than 2, formula often 5-5-4 in male; front coxae small, rounded; species strongly flattened, parallel-sided; antennae often long and more or less filiform; head more or less sharply contracted behind temples (fig. 89)
	temples (fig. 89)
11	Front coxae rounded or somewhat ovate, their cavities closed behind; elytral epipleurae distinct and complete; tarsal formula 5-5-5 in both sexes, more or less pseudotetramerous; upper surface nearly always glabrous; antennal club strong9. EROTYLIDAE
	Front coxae more transverse, their cavities open behind
12	Front coxal cavities nearly closed behind; tarsal formula 5-5-5 in both sexes; elytra somewhat truncate, not completely covering last abdominal tergite (fig. 94); prothorax practically as long as wide; species less than 1.5 mm. long; antennal insertions widely separated
	(Hypocoprus Mots., H. quadricollis Reitt.) Front coxal cavities widely open behind; tarsi usually 5-5-4 in male; prothorax
	transverse; elytra not truncate, completely covering abdomen; if length less than 1.5 mm., antennal inserted approximations8. CRYPTOPHAGIDAE

13	Species very minute (length not more than 1mm.); form broad and smoothly
	ovate, not very convex (fig. 95), prothorax more or less semicircular in outline
	usually hiding head; antennal segments 1-2 much wider than 3-5; usually
	six visible abdominal sternites
	If species not more than 1 mm. long, shape different14
14	Front coxae distinctly transverse; tarsal formula 4-4-4 or 3-4-4, without lobed
	segments; form more or less oblong, usually pubescent
	If front coxae distinctly transverse, tarsi pseudotrimerous
15	Prothorax of Bostrychoid type (cf. fig. 12); antennae 8-10 segmented with loose
	normal 3-segmented club; elytra without regular striae; trochanters normal
	(cf. fig. 19)
	Prothorax not of Bostrychoid shape; antennae 11-segmented with 3-4 segmented
	club; trochanters heteromeroid (fig. 21); tarsal formula 3-4-4 in male
	20. Mycetophagidae
16	Tarsi 4-4-4, segment 2 not lobed below; front coxae small, rounded; elytra
	usually more or less striate
	Tarsi more or less pseudotrimerous or 3–3–3 segmented



96
FIGS. 93-98.—93, Laria dulcamarae Scop. 94, Hypocoprus quadricollis Reitt. 95
Sericoderus lateralis Gyll. 96, Anommatus 12-striatus Muell. 97, Holoparamecus caularum Aubé. 98, Aderus populneus Panz.



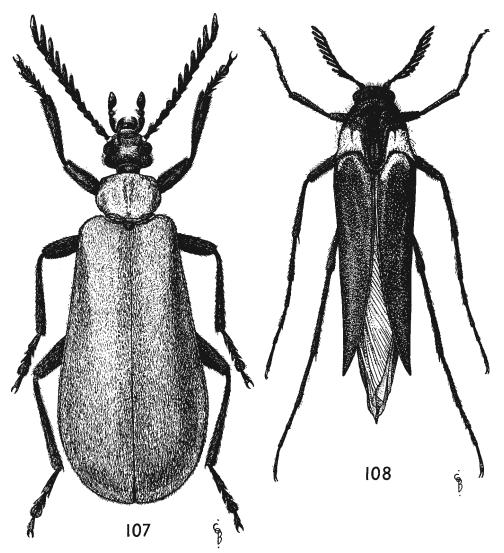
Figs. 99–102.—99, Biphyllus lunatus F. 100, Byturus tomentosus Deg. 101, Tetratoma fungorum F. 102, Lagria hirta L.

23. LAGRIIDAE

- Apical segment of maxillary palpi at least as large as penultimate, not pointed at apex; trochanters more or less heteromeroid (fig. 21), sometimes scarcely visible in ventral view; antennal insertions often hidden under sides of front; if general form broad, dorsal surface rough, dull and more or less pubescent 19. COLYDIIDAE Apical segment of maxillary palpi smaller than penultimate, pointed at apex (cf. fig. 1L); trochanters more or less long (cf. fig. 20); antennal insertions not hidden under sides of front; form rather short and broad, upper surface glabrous and Tarsi pseudotrimerous; front coxae more or less transverse; form short, ovate Tarsal formula 3-3-3, segment 2 not lobed below; front coxae small, rounded; Last segment of maxillary palpi not or scarcely securiform; antennae not very short; metasternum and first visible abdominal sternite without femoral lines; Upper surface smooth, glabrous (figs. 96, 97); middle coxal cavities closed outwardly by sterna; prosternal process relatively broad; upper surface of head smooth, clypeus in same plane as frons, separated from it by a normal suture MEROPHYSIIDAE Upper surface more or less rugose or strongly punctured or closely pubescent; middle coxal cavities not closed outwardly by sterna; upper surface of head more or less uneven, clypeus not in the same plane as frons...17. LATHRIDIIDAE First three visible abdominal sternites connate (cf. fig. 17); front coxal cavities Abdomen with not more than two basal sternites connate; front coxal cavities not visibly closed behind......24 Elytra with long outstanding pubescence; prothorax without distinct side borders (fig. 102); front coxae projecting, prosternal process evanescent between them
  - 103 105 106

Figs. 103-106.—103, Sphaeriestes castaneus Panz., head and prothorax, dorsal view. 104, Mordella aculeata L., outline of body, dorsal view. 105, Mordellid tarsal claws. 106, Alleculid tarsal claw. AP, appendages below claws; C, claw.

Elytra without long outstanding pubescence; side borders of prothorax usually well-marked, its base not or little narrower than elytra at shoulders; front coxae not distinctly projecting, prosternal process easily visible between them

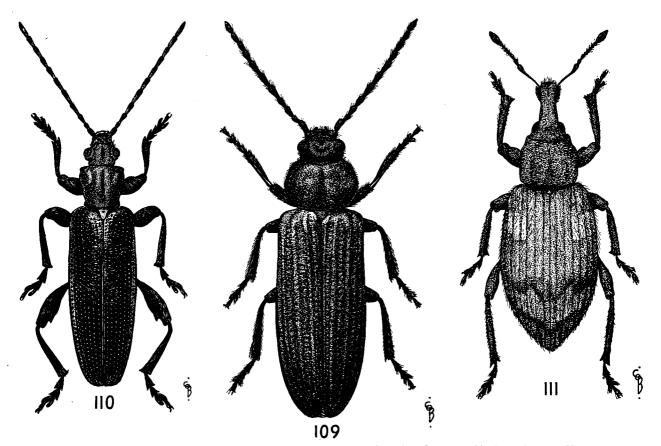


Figs. 107-108.—107, Pyrochroa coccinea L. 108, Metoecus paradoxus L.

24	Prothorax with side-edges distinct, at least in basal half, its basal margin nearly or quite as wide as elytra at shoulders; antennae never flabellate
25	Prothorax without distinct side edges
26	Antennae without a strong 4-segmented club; general form more or less navicular (cf. fig. 104)
27	Head with a narrow neck, its posterior face excavate to fit closely against prothorax; antennae never clubbed
28	Last abdominal tergite not produced into a spine; outer face of hind tibiae without such spinules
29	lobed
30	Head not rostrate; if two basal abdominal sternites connate, head with a narrow neck
31	Form less flattened; antennae longer; head with a narrow neck, or penultimate tarsal segment lobed below
<b>3</b> 2	if elytra not meeting along suture, head with broad neck or wings vestigial32 Head not strongly deflexed, neck relatively broad (fig. 107)
33	Head strongly deflexed, neck very narrow (fig. 98)
34	general form narrower, less flattened
35	segment more or less lobed, without appendages below claws; elytra always fully covering abdomen

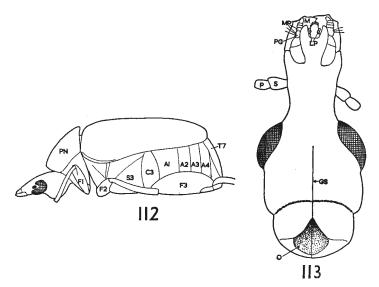
## Superfamily 17.—CHRYSOMELOIDEA.

Any species with a really definite antennal club can be excluded from this group and sought under Cleroidea, Cucujoidea or Curculionoidea. Some SILVANIDAE (Cucujoidea) which might be sought here can be distinguished by their small size, testaceous to fuscous colouration, and dentate side of



Figs. 109-111,-109, Asemum striatum L. 110, Plateumaris sericea L. 111, Alophus triguttatus F.

the prothorax; certain CLERIDAE (Cleroidea) with Chrysomeloid-like tarsi and antennae are distinguished by their outstanding bristles, projecting front coxae, and six visible abdominal sternites.



Figs. 112-113.—112, Bruchus rufimanus Boh., outline of body, lateral view. 113, Deporaus betulae L., head, ventral view. A1, A2, A3, A4, visible abdominal sternites 1-4; C3, hind coxa; F1, F2, F3, front, middle and hind femora; GS, fused gular sutures; LP, labial palp; M, mandible; MP, maxillary palp; P, pedicel; PG, palpiger; PN, pronotum; S, scape; S3, metasternum; O, occipital foramen; T7, last visible abdominal tergite.

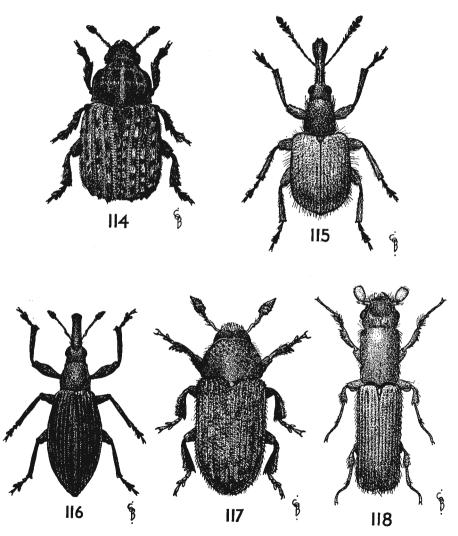
## Superfamily 18.—CURCULIONOIDEA.

Some approach to the Curculionoid rostrum is met with in the Cucujoid Salpingidae and Mycteridae, and to a less degree in the Chrysomeloid Bruchidae; conversely a weak or non-existent rostrum is seen in the Curculionoid Scolytidae and Platypodidae, also some Anthribidae.

The Scolytids and Platypodids, also certain Curculionids, show reduction or loss of the lobing of the third segment.

1 Rostrum very short; antennae not geniculate, with loose 3-segmented club; prothorax with a basal or pre-basal transverse keel which extends forwards on each side forming a more or less imperfect side-border (fig. 114); four basal abdominal sternites connate; labrum distinct; maxillary palpi exserted, flexible; tarsal segment 3 more or less deeply enclosed in lobes of 2

2. Anthribidae



Figs. 114-118.—114, Brachytarsus fasciatus Först. 115, Caenorhinus aequatus L. 116, Apion radiolus Kirby. 117, Hylesinus fraxini Panz. 118, Platypus cylindrus F.

If rostrum very short, antennae geniculate and with a compact club; if prothorax Labrum free; maxillary palpi exserted, flexible; all abdominal sternites free; antennae not at all geniculate, with a very weak 3-segmented club; rostrum (Cimberis Gozis., C. attelaboides F.) Labrum not distinct; maxillary palpi concealed, rigid; at least two basal abdominal sternites connate; antennal club more compact; elytra nearly always striate......3 Antennae straight, club very distinctly segmented (fig. 115); tarsal claws connate or mandibles toothed on outer edges (fig. 113); upper surface shining, often glabrous, never with scales; if trochanters long, rostrum very short ATTELABIDAE Antennal club compact; if antennae straight, tarsal claws free and mandibles not toothed on outer edge; upper surface rarely so shining, often with scales.....4 Rostrum more or less distinct; tibiae never with dentate outer edges, prothorax Rostrum vestigial or absent (figs. 117-118); tibiae with dentate outer edges or prothorax with distinct side borders......6 Trochanters long; antennae straight or prothorax at base almost as wide as elytra at shoulders; rostrum more or less long (fig. 116)..............4. APIONIDAE Trochanters normal; antennae nearly always geniculate; base of prothorax much narrower than elytra at shoulders (fig. 111)..........5. CURCULIONIDAE Tarsi with segment 4 very small; general shape less elongate and cylindrical Tarsi with segment 4 at least half as long as 3; shape elongate and cylindrical 

Superfamily 19.—STYLOPOIDEA.
With a single family, STYLOPIDAE

#### CONSPECTUS OF THE CLASSIFICATION ADOPTED IN THIS PART.

Cross-referenced to the equivalent groupings in Fowler's Coleoptera of the British Islands (followed by Beare's Catalogue of the Coleoptera of the British Isles), Reitter's Fauna Germanica: Käfer, Imms' General Textbook of Entomology, Joy's Practical Handbook of British Beetles, van Emden's Larvae of British Beetles (1939–1949), and Kloet and Hincks' Check list of British Insects. Where not specifically cited, a work may be assumed to use the same name for the same grouping as this Part, or (in the case of Imms) to make no reference to the group at all.

Suborder ADEPHAGA (Geodephaga + Hydradephaga + Gyrinia Joy)

Superfamily Caraboidea (Geodephaga + Hydradephaga + Gyrinia Joy; Caraboidea + Hydradephaga Kloet & Hincks).

- CARABIDAE (Cicindelidae + Carabidae Fowler, Reitter, Joy, Imms, van Emden).
- 2. Haliplidae.
- 3. Hygrobiidae (Pelobiidae Fowler, Imms).
- NOTERIDAE (Dytiscidae—Noterina Fowler; -Noterina Reitter; -Noterinae Joy, van Emden, Kloet & Hincks).
- 5. Dytiscidae auctt, minus Noterus).
- 6. Gyrinidae.

#### Suborder POLYPHAGA.

Superfamily 1.—Sphaerioidea (Clavicornia pars Fowler, Joy; Staphylinoidea pars Reitter, Imms, van Emden, Kloet & Hincks).

#### 1. SPHAERIIDAE.

Superfamily 2.—**Hydrophiloidea** (Clavicornia pars Fowler; Diversicornia pars Imms; Palpicornia + Georyssidae Reitter, Joy, Kloet & Hincks; Hydrophiloidea van Emden minus Histeridae).

- Hydraenidae (Hydrophilidae-Hydrochina minus Hydrochus + Limnebius Fowler; -Hydraeninae minus Hydrochus + Limnebius Reitter; -Hydraeninae + Limnebiinae Kloet & Hincks; Hydraenidae-Hydraeninae minus Hydrochus + Hydrophilidae-Limnebiinae Joy; Hydrophilidae pars Imms, van Emden).
- 2. Spercheidae (Hydrophilidae-Spercheina Fowler; -Spercheinae Reitter, Kloet & Hincks, van Emden; Hydraenidae-Spercheinae Joy).

- 3. Hydrochidae (Hydrophilidae-Hydrochinae Hydrochus Fowler; -Hydraeninae Hydrochus Reitter; -Hydrochinae Kloet & Hincks, van Emden; Hydraenidae-Hydraeninae Hydrochus Joy).
- 4. Georissidae (Georyssidae Fowler, Reitter, Joy, Imms).
- 5. HYDROPHILIDAE (Hydrophilidae auctt. minus Hydraeninae, Limnebiinae, Spercheinae, Hydrochinae).
- Superfamily 3.—Histeroidea (Clavicornia pars Fowler, Joy; Necrophaga pars + Histerida pars Reitter; Staphylinoidea-Histeridae + Diversicornia-Sphaeritidae Imms; Staphylinoidea-Histeridae plus Claviconia-Sphaeritidae Kloet & Hincks; Hydrophiloidea-Histeridae van Emden).

—. SPHAERITIDAE (Silphidae-Sphaeritina Fowler;
 -Sphaeritini Reitter).

2. HISTERIDAE.

- Superfamily 4.—Staphylinoidea (Clavicornia pars Fowler; Staphylinoidea minus Clambidae, Corylophidae, Sphaeridae, Hydroscaphidae, Silphidae-Sphaeritini, Histeridae Reitter; Staphylinoidea minus Hydroscaphidae, Sphaeriidae, Corylophidae, Histeridae, Silphidae-Clambinae Imms; Brachelytra + Clavicornia pars Joy; Staphylinoidea minus Clambidae, Orthoperidae, Sphaeriidae, Histeridae Kloet & Hincks).
  - 1. PTILIDAE (Trichopterygidae Fowler, Imms, van Emden).
  - 2. LEPTINIDAE (Silphidae Leptininae Kloet & Hincks).
  - 3. Anisotomidae (Silphidae-Anisotomina + Cholevinae Fowler; Silphidae-Cholevinae + Liodidae Reitter; Silphidae pars Imms, van Emden; Anisotomidae + Cholevidae Joy; Silphidae-Bathysciinae + Catopinae + Coloninae + Leiodidae Kloet & Hincks).
  - 4. SCYDMAENIDAE.
  - 5. SCAPHIDIIDAE.
  - 6. MICROPEPLIDAE (Staphylinidae-Micropeplinae Reitter, Imms, Joy; Staphylinidae-Oxytelinae *Micropeplus* Kloet & Hincks).
  - 7. STAPHYLINIDAE (Staphylinidae minus *Micropeplus* Reitter, Imms, Joy, Kloet & Hincks).
  - 8. PSELAPHIDAE.

Superfamily 5.—Scarabaeoidea (Lamellicornia Fowler, Reitter, Imms, Joy, van Emden, Kloet & Hincks).

- 1. LUCANIDAE
- 2. Trogidae (Scarabaeidae Laparosticti Trogina Fowler; Scarabaeidae-Coprophaginae-Trogini Reitter; Scarabaeidae-Coprophaginae *Trox* Joy; Scarabaeidae Troginae Imms, van Emden, Kloet & Hincks).
- 3. Geotrupidae (Scarabaeidae Laparosticti Geotrupina Fowler; Scarabeidae Coprophaginae-Geotrupini Reitter; Scarabaeidae Coprophaginae Odontaeus + Ceratophyus + Geotrupes Joy; Scarabaeidae Geotrupinae Imms, van Emden, Kloet & Hincks).
- 4. SCARABAEIDAE (Scarabaeidae auctt. minus Troginae, Geotrupinae).

Superfamily 6.—Dascilloidea (Malacodermata-Dascillidae minus Eubria + Silphidae-Clambina Fowler; Malacodermata pars + Necrophaga-Clambidae Reitter; Diversicornia pars + Staphylinoidea-Clambinae Imms; Malacodermata pars + Clavicornia-Clambidae Joy; Fossipedes minus Eubria + Staphylinoidea-Clambidae Kloet & Hincks; Dascilloidea-Dascillidae + -Helodidae + Staphylinoidea-Clambidae van Emden).

- 1. CLAMBIDAE (Silphidae-Clambina Fowler).
- 2. HELODIDAE (Dascillidae-Cyphonina minus *Eubria*Fowler; Helodidae-Helodinae Reitter; Helodidae minus *Eubria* Joy; Helodidae pars Imms).
- 3. Dascillidae (Dascillidae Dascillina Fowler; Dascillidae pars Imms; Dascillidae minus Eubria Kloet & Hincks).
- Superfamily 7.—**Byrrhoidea** (Clavicornia pars Fowler, Joy; Brachymera pars Reitter, Kloet & Hincks; Diversicornia pars Imms; Dascilloidea-Byrrhidae van Emden).
  - 1. Byrrhidae minus Aspidiphorus and Limnichus Fowler; -Byrrhini Reitter; Byrrhidae pars Imms; Byrrhidae minus Limnichus Joy, Kloet and Hincks).
- Superfamily 8.—**Dryopoidea** (Malacodermata-Eubria + Clavicornia-Heteroceridae, Parnidae and Limnichus Fowler; Malacodermata-Eubriinae + Brachymera-Limnichini + Hygrophili minus Georyssidae Reitter; Malacodermata-Eubria + Clavicornia-Parnidae, Helmidae, Heteroceridae and Limnichus Joy; Diversicornia pars Imms; Macrodactylia minus Georissidae + Fossipedes-Eubria + Brachymera-Limnichus Kloet & Hincks; Dascilloidea-Dryopidae + Heteroceridae van Emden).

- 1. PSEPHENIDAE (Dascillidae Cyphonina Eubria Fowler; Helodidae-Eubrinae Reitter; Helodidae-Eubria Joy; Dryopidae pars Imms; Dascillidae-Eubria Kloet & Hincks).
- 2. HETEROCERIDAE.

3. LIMNICHIDAE (Byrrhidae-*Limnichus* Fowler, Joy, Kloet & Hincks; -Limnichini Reitter).

- 4. Dryopidae (Parnidae-Parnia Fowler; Dryopidae-Dryopini Reitter; Dryopidae pars Imms; Parnidae Joy; Dryopidae-Dryopinae Kloet & Hincks).
- 5. ELMIDAE (Parnidae-Elmina Fowler; Dryopidae-Potamophilini + Helminthinae Reitter; Helmidae Joy; Dryopidae pars Imms; Dryopidae-Elminae Kloet & Hincks).
- Superfamily 9.—Buprestoidea (Sternoxi pars Fowler; Sternoxia pars Reitter, Joy, Kloet & Hincks; Diversicornia pars Imms; Sternoxia-Malacodermata pars van Emden).
  - 1. Buprestidae.

Superfamily 10.—**Elateroidea** (Sternoxi pars Fowler; Sternoxia pars Reitter, Joy, Kloet & Hincks; Diversicornia pars Imms; Sternoxia-Malacodermata pars van Emden).

Elateridae (Elateridae-Elaterinae Imms; Elateridae minus Trixagus, Melasis, Dirhagus and

Eucnemis Joy).

2. TRIXAGIDAE (Throscidae Fowler, Imms; Ela-

teridae-Trixagus Joy).

3. EUCNEMIDAE (Elateridae - Eucneminae Imms; Elateridae genn. *Melasis*, *Dirhagus* and *Eucnemis* Joy).

- Superfamily 11.—Cantharoidea (Malacodermata Cantharidae minus Malachiinae Reitter; Diversicornia pars Imms; Malacodermata pars Fowler, Joy, Kloet & Hincks; Malacodermata-Sternoxia pars van Emden).
  - DRILIDAE (Cantharidae-Drilinae Reitter, Imms; Cleridae-Drilus Joy; Telephoridae-Drilinae van Emden).

2. Lampyridae (Cantharidae-Lampyrinae Reitter, Imms; Telephoridae-Lampyrinae van Emden)

- 3. CANTHARIDAE (Telephoridae Fowler; Cantharidae-Cantharinae Reitter, Imms; Cantharidae pars Joy; Telephoridae-Telephorinae van Emden).
- 4. Lycidae (Cantharidae-Lycinae Reitter, Imms; Telephoridae-Lycinae van Emden).

- Superfamily 12.—Dermestoidea (Clavicornia pars Fowler, Joy; Brachymera pars Reitter, Kloet & Hincks; Diversicornia pars Imms; Dascilloidea-Dermestidae + Teredilia-Thorictidae van Emden).
  - 1. THORICTIDAE.
  - 2. Dermestidae.
- Superfamily 13.—Bostrychoidea (Ptinoidea minus Sphindidae, Cisidae Fowler; Teredilia + Clavicornia-Lyctidae Reitter; Diversicornia pars Imms; Teredilia minus Orthoceridae + Clavicornia-Lyctidae Joy; Teredilia Kloet & Hincks; Teredilia minus Cisidae and Mordellidae van Emden).
  - Anobiidae (Anobiidae + Ptinidae-Hedobia Fowler; Anobiidae-Anobiides Imms; Anobiidae + Dorcatomidae + Xyletinidae + Ptinidae-Hedobia Joy).
  - PTINIDAE (Ptinidae minus Hedobia Fowler, Joy; Anobiidae-Ptinides Imms).
  - 3. Bostrychidae (Bostrichidae Fowler, Imms, Kloet & Hincks).
  - 4. LYCTIDAE.
- Superfamily 14.—Cleroidea (Clavicornia-Trogositidae + Malacodermata pars Fowler, Reitter, Kloet & Hincks; Diversicornia pars Imms; Clavicornia pars + Malacodermata pars Joy; Sternoxia-Malacodermata-Melyridae, Trogositidae and Cleridae van Emden).
  - Trogositidae (Ostomidae Reitter; Ostomatidae Kloet & Hincks; Cucujidae-Trogositinae + Colydiidae-Nemosoma + Mycetophagidae-Thymalinae Joy).

2. CLERIDAE (Cleridae Joy minus Drilus).

- 3. MELYRIDAE (Melyridae minus Phloeophilina Fowler; Cantharidae-Malachiinae + Dasytinae minus Phloeophilus Reitter; Melydridae Imms; Cantharidae pars Joy; Malachiidae + Dasytidae minus Phloiophilus Kloet & Hincks).
- 4. Phloiophilidae (Melyridae-Phloeophilina Fowler; Cantharidae-Dasytinae-Phloeophilus Reitter; Mycetophagidae Mycetophaginae Phloeophilus Joy; Dasytidae Phloiophilus Kloet & Hincks).
- Superfamily 15.—Lymexyloidea (Malacodermata pars Fowler, Reitter, Joy, Kloet & Hincks; Diversicornia pars Imms; Clavicornia-Heteromera pars van Emden).
  - LYMEXYLIDAE (Lymexylonidae Fowler, Reitter, Imms. Joy).

Superfamily 16.—Cucujoidea (Clavicornia pars + Ptinoidea pars + Heteromera Fowler; Staphylinoidea-Corylophidae + Clavicornia minus Ostomidae and Lyctidae + Heteromera Reitter; Staphylinoidea-Corylophidae + Diversicornia pars + Heteromera Imms; Heteromera and Teredilia-Orthoceridae + Clavicornia pars Joy; Staphylinoidea-Orthoperidae + Clavicornia minus Sphaeritidae and Ostomatidae + Heteromera Kloet & Hincks; Clavicornia-Heteromera minus Lymexylidae + Teredilia-Cisidae and Mordellidae van Emden).

1. NITIDULIDAE (Nitidulidae minus Rhizophagina Fowler; Nitidulidae minus Rhizophaginae Reitter, Imms; Nitidulidae-Nitidulinae + -Erotylinae-Glischrochilus + Cateretidae + Colydiidae-Pityophagus Joy).

2. Rhizophagidae (Monotomidae + Nitidulidae-Rhizophaginae Fowler, Reitter, Imms; Monotomidae + Colydiidae-Rhizophagus Joy; Rhizophagidae + Cucujidae-Monotoma Kloet & Hincks; Monotomidae + Rhizophagidae van Emden).

3. PHALACRIDAE.

4. CUCUJIDAE (Cucujidae-Cucujina + -Hyliotina Fowler; Cucujidae-Cucujinae minus Silvanini, Psammoechini, Prostomini Reitter; Cucujidae pars Imms, van Emden; Cucujidae-Uleiotinae + Cucujinae Joy; Cucujidae genn. Uleiota, Pediacus, Dendrophagus and Laemo-phloeus Kloet & Hincks).

5. SILVANIDAE (Cucujidae-Psammoechina + -Silvalina Fowler; -Silvanini + -Psammoechini Reitter; Cucujidae pars Imms, van Emden; Cucujidae-Psammoecinae + Cryptophagidae-Silvaninae Joy; Cucujidae genn. Nausibius, Ahasuerus, Oryzaephilus, Silvanus, Silvanoprus, Cryptamorpha and Psammoecus Kloet & Hincks).

6. Sphindidae + Byrrhidae-Aspidiphorus Fowler; Sphindidae+Mycetophagidae-Aspidiphorinae Joy; Sphindidae + Aspidiphoridae Kloet & Hincks).

7. Hypocopridae (Cucujidae-Hypocoprinae Fowler; Cryptophagidae-Hypocoprini Reitter; Lathrididae-Hypocoprus Joy; Cucujidae-Hypocoprus Kloet & Hincks).

8. CRYPTOPHAGIDAE (Cryptophagidae minus Diphyllina Fowler; Cryptophagidae-Telmatophilini minus Cryptophilus + -Cryptophagini minus Pharaxonotha + Atomariini Reitter; Cryptophagidae pars Imms; Cryptophagidae

 $\begin{array}{lll} {\rm Cryptophaginae\ minus\ } Diplocoelus + {\rm Myceto-} \\ {\rm phagidae-Atomariinae\ } & + {\rm Lathridiidae-} \\ {\it Paramecosoma\ and\ } Telmatophilus + {\rm Mycetophagi-} \\ \end{array}$ 

dae Antherophagus Joy).

9. EROTYLIDAE (Erotylidae minus Languriinae Imms; Nitidulidae-Erotylinae genn. Tritoma, Triplax and Dacne Joy; Erotylidae minus Biphyllus and Diplocoelus Kloet & Hincks; Erotylidae minus Diphyllus and Tetratoma van Emden).

10. BIPHYLLIDAE (Cryptophagidae-Diphyllina Fowler; -Diphyllini Reitter; Cryptophagidae-Cryptophaginae-Diphocoelus + Nitidulidae-Erotylinae-Diphyllus Joy; Erotylidae-Biphyllus + Diplocoelus Kloet & Hincks; Erotylidae-Diphyllus van Emden).

11. BYTURIDAE (Mycetophagidae-Byturinae Joy).

- 12. Endomychidae (Endomychidae + Lathridiidae-Mycetaea + Mycetophagidae-Sphaerosominae Joy; Endomychidae minus Murmidiinae van Emden).
- 13. Coccinellidae.

14. Corylophidae (Orthoperidae Kloet & Hincks).

15. CERYLONIDAE (Colydiidae-Cerylonina + -Murmidiina Fowler; -Ceryloninae-Cerylonini + -Murmidiinae Reitter; Colydiidae pars Imms; Colydiidae-Cerylon Joy; Colydiidae-Ceryloninae-Cerylon + -Murmidiinae Kloet & Hincks; Endomychidae - Murmidiinae van Emden).

16. Merophysiidae (Lathridiidae-Merophysiina Fowler; Colydiidae-Ceryloninae-Anommatini + Lathridiidae - Holoparamecini Reitter; Colydiidae-Anommatus + Lathridiidae-Holoparamecus Joy; Lathridiidae-Holoparamecinae + Colydiidae - Ceryloninae - Anommatus Kloet & Hincks).

17. LATHRIDHDAE (Lathridiidae minus Merophysiina Fowler; -Lathridiini + -Corticariini Reitter; Lathridiidae minus Mycetaea, Myrmecoxenus, Ditoma, Hypocoprus, Paramecosoma, Telmatophilus and Holoparamecus Joy; -Lathridiinae Kloet & Hincks; Lathridiidae minus Holoparamecus van Emden).

18. CISIDAE (Cisidae Fowler; Cioidae Imms; Ciidae

Kloet & Hincks).

19. COLYDIDAE (Colydiidae-Deretaphrina + Synchitina + Langelandiina Fowler; Colydiidae-Colydiinae + -Ceryloninae-Bothriderini and -Deretaphrini Reitter; Colydiidae pars Imms; Orthoceridae+Lathridiidae-Ditoma and -Myr-

mecoxenus + Colydiidae minus Anommutus, Cerylon, Nemosoma, Pityophagus and Rhizophagus Joy; Colydiidae-Colydiinae + -Cryloninae-Teredus and -Oxylaemus Kloet & Hincks; Colydiidae minus Cerylon van Emden)

20. Mycetophagidae - Mycetophagidae - Mycetophaginae minus Antherophagus and Phloeo-

philus Joy).

21. Tetratomidae (Melandryidae - Tetratomina Fowler; Tetratomini Reitter; MelandryidaeTetratoma Joy; Serropalpidae-Tetratoma Kloet & Hincks; Erotylidae-Tetratoma van Emden).

22. TENEBRIONIDAE.

23. LAGRIIDAE.

24. Alleculidae (Cistelidae Fowler, Imms, Joy).

25. Mycteridae (Pythidae - Mycterina Fowler; -Mycterini Reitter; -Mycterus Kloet & Hincks).

26. Salpingidae (Pythidae-Salpingina Fowler; -Salpingini Reitter; Pythidae pars Imms; Pythidae minus Pytho Joy; Pythidae minus Pytho and Myterus Kloet & Hincks).

27. PYTHIDAE (Pythidae-Pythina Fowler; -Pythini Reitter; Pythidae pars Imms; Pythidae-

Pytho Joy, Kloet & Hincks).

28. PYROCHROIDAE.

29. OEDEMERIDAE.

30. Melandryidae (Melandryidae Melandryina Fowler; Melandryidae minus Tetratomini, Stenotrachelini and Hallomenini - Mycetoma Reitter; Melandryidae pars Imms; Melandryidae minus Tetratoma Joy; Serropalpidae minus Tetratoma Kloet & Hincks; Melandryidae + Synchroidae van Emden).

31. SCRAPTHDAE (Scraptiidae + Mordellidae - Anaspina Fowler; Mordellidae-Scraptiinae + -Mordellinae-Anaspini Reitter; Mordellidae-Scraptiinae + -Mordellinae-Anaspis Joy; Scraptiidae + Mordellidae-Anaspis Imms, van Emden, Kloet & Hincks).

32. Mordellidae (Mordellidae-Mordellina Fowler; -Mordellini Reitter; -Mordellinae minus Anaspis Joy; Mordellidae minus Anaspis Imms, van Emden, Kloet & Hincks).

- 33. Rhipiphoridae (Rhipidophoridae Fowler).
- 34. MELOIDAE.
- 35. ANTHICIDAE (Anthicidae-Anthicinae Reitter).

 ADERIDAE (Xylophilidae Fowler, Imms, Kloet & Hincks; Anthicidae-Hylophilinae Reitter; van Emden; Hylophilidae Joy).

Superfamily 17.—Chrysomeloidea (Longicornia + Phytophaga Fowler, Joy; Phytophaga Reitter, Imms, van Emden, Kloet & Hincks).

CERAMBYCIDAE (Prionidae + Cerambycidae + Lamiidae Fowler; Prionidae + Cerambycidae + Tetropiidae + Lamiidae Joy).

2. Bruchidae (Lariidae Reitter, Joy).

3. Chrysomelidae (Cryptostomidae + Chrysomelidae Joy).

Superfamily 18.—Curculionoidea (Rhynchophora Fowler, Reitter, Imms, van Emden, Kloet & Hincks; Rhynchophora + Clavicornia-Choragidae Joy).

1. Nemonychidae (Rhinomaceridae Fowler; Curculionidae-Rhynchitinae-Nemonychini Reitter; Curculionidae pars Imms; Curculionidae-Rhynchitinae-Nemonychini Reitter; Curculionidae pars Imms; Curculionidae-Rhynchitinae-Rhinomacer Joy, Kloet & Hincks).

2. Anthribidae (Platyrrhinidae Fowler; Anthribidae + Choragidae Joy; Platystomidae Kloet

& Hincks).

3. ATTELABIDAE (Curculionidae-Attelabinae Fowler; -Rhynchitinae minus Nemonychini Reitter; Curculionidae pars Imms, van Emden; -Rhynchitinae minus Rhinomacer Joy; -Rhynchitinae minus Rhinomacer + Attelabinae + Apoderinae Kloet & Hincks).

APIONIDAE (Curculionidae-Apioninae + -Curculioninae-Cionina-Nanophyes Fowler; -Apioninae Reitter; -Apioninae + -Nanophyinae Joy; Curculionidae pars Imms, van Emden; -Apioninae + -Mecininae-Nanophyes Kloet & Hincks).

CURCULIONIDAE (Curculionidae-Brachyrrhininae
 + -Curculioninae minus Nanophyes Fowler;
 -Adelognatha +- Phanerognatha minus Apioninae
 and Rhynchitinae Reitter; Curculionidae
 minus Rhynchitinae, Apioninae and Nanophyinae Joy; Curculionidae pars Imms, van
 Emden; Curculionidae minus Rhynchitinae,
 Attelabinae, Apoderinae, Apioninae and Mecininae-Nanophyes Kloet & Hincks).

 SCOLYTIDAE (Ipidae Reitter; Scolytidae + Platypodidae Kloet & Hincks, van Emden).
 (Note: in a strictly natural classification the last

two families should probably be merged.)

- Superfamily 19.—Stylopoidea (Abnormal Coleoptera Fowler; not treated by Reitter or Joy; Order Strepsiptera Imms, Kloet & Hincks; Clavicornia-Heteromera-Stylopidae van Emden).
  - 1. STYLOPIDAE.

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